

# **Food and Food Packaging**

It was recognized fairly early that PFAS compounds used in food packaging materials (such as pizza boxes and microwave popcorn bags) could migrate into consumable food products and contribute to increased PFAS body burden. In addition, as PFAS contamination has continued to spread throughout the environment it has been more recently recognized that these materials can also enter the human food supply chain through animal consumption of PFAS contaminated water and feed, thereby further increasing our PFAS body burden. Regardless of source, the analysis of PFAS in food and food packaging materials - and their myriad complex matrices - features additional difficult analytical challenges.



## 1. New Concerns about PFAS in Food

### The Convergence of Environmental Contamination and Food Safety

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#### **Abstract**

Per- and Polyfluorinated Alkyl Substances (PFAS) are well known environmental contaminants that have a newly recognized potential to taint certain food products through agricultural consumption via environmental transport from contaminated industrial sites [1]. The analysis of PFAS in food products requires more extensive analytical preparation techniques, compared to PFAS testing of simple matrices such as drinking water, in order to reduce the impact of sample matrix interferences on the subsequent instrumental analysis. An example is provided of a PFAS method applicable to milk, butter, cheese and fish.

### The Prequel

Per- and Polyfluorinated Alkyl Substances (PFAS) are an extensive family of synthetic, fluorochemicals with a unique set of physical and chemical properties. These properties have resulted in their widespread commercial use over the past 50 years in diverse applications ranging from fire fighting foams to stain resistant carpet to grease-proof pizza boxes. However, these same unique physical and chemical properties also have been found to bear serious environmental consequences: widespread dispersion ability, extreme environmental persistence and a high degree of bioaccumulation [2]. Although PFAS do not exhibit acute toxic properties, researchers have found that PFAS can demonstrate a large number of subtle, chronic health effects, primarily affecting the endochrine and reproductive systems. Consequently, health experts have long been concerned that low-level, cumulative exposure to PFAS over an extended period of time could have serious health consequences [3]. Therefore, chronic lifetime PFAS exposure pathways - such as through food or drinking water - are of particular concern to regulators and are receiving enhanced scrutiny.

### **Initial Concerns**

In the US, the initial US Food and Drug Administration (FDA) concern about PFAS centered about the contamination of food products through contact with PFAS containing food packaging (and to a lesser extent with food processing equipment). The classic examples are those PFAS coated pizza boxes, fast-food hamburger wrappers and microwave popcorn bags that have done such a marvelous job of keeping grease off our clothes. That problem was summarily solved in late 2016 when FDA removed the approval for the use of PFAS in food packaging [4].

Likewise, the primary US Environmental Protection Agency (EPA) focus has been on drinking water as a primary source of lifetime PFAS exposure. EPA is continuing to conduct extensive nationwide testing for PFAS in drinking water under the Unregulated Contaminant Monitoring Rule (UCMR) program [5]. These efforts will very likely result in specific regulatory limits for the allowable concentration of certain PFAS in drinking water.

Concurrently, other government agencies, such as the US Department of Defense (DOD) have been extensively studying the widespread environmental contamination of military facilities owing to the extensive historical use of PFAS firefighting foams, principally at air bases [6].

### Convergence

Initially, these three individual trains of concern seemed to be running on separate tracks. It was only more recently that they were seen to be converging toward a much larger, more complex problem requiring multimedia, multi agency examination and the use of more sophisticated analytical tools. The simplified pathway model shown in **Figure 1** illustrates the general scope of the problem. By the end of 2019, the FDA was fully on board with concerns about PFAS entering the general food supply through environmental sources, potentially leading to the contamination of dairy products, bottled water, seafood and other consumables [7].

### **Analytical Implications**

This expanded concept of the PFAS problem is clearly a major step forward, but it has presented some analytical challenges. Much of the official PFAS methodology developed over the past decade has been focused on the analysis of drinking water and aimed at a very limited list of analytes. With little challenge from matrix interference, easily surmountable chromatography issues and straight forward mass spectrometry, these official drinking-water-only methods proved to be inadequate when applied to the analysis of PFAS in soil, sediment, sludge and wastewater. When applied to the analysis of foods - with a myriad of complex matrices, they are quite ineffective, resulting in a surge in PFAS analytical method development centered about complex matrices, with food testing occupying a prominent position. The following section features one such application as an illustration of the approaches now being pursued in pursuit of the expanded PFAS challenge.

## Analysis of PFAS in Dairy Products, Eggs, and Fish by LC-MS/MS

### **Method Introduction**

The following work was performed through a collaboration between Weck Laboratories, Inc., City of Industry, CA, USA and Phenomenex, Inc., Torrance, CA, USA, for the development of new sample preparation and analysis procedures for determining low levels of PFAS in food products. This particular application was directed at achieving sub-ppb sensitivity for 23 PFAS analytes in dairy products (milk, butter and cheese), eggs and fish as representative of difficult to analyze fatty matrices. The following discussion is a synopsis of the full work [8].

### **Sample Preparation**

One gram of homogenized sample was spiked with internal standards and surrogates and an analyte mix of 23 PFAS compounds (**Table 1**) at the 1 ng/g level, followed by the addition of 10 mL acetonitrile and 10 mL water. Four replicates of each matrix (milk, eggs, butter, cheese and fish) were prepared. The samples were processed by a modified QuEChERs procedure using a commercial kit (Phenomenex roQ™ Extraction Kit). An aliquot (500 µL) of the cleaned acetonitrile phase was transferred to an LC vial for analysis. **Figure 2** displays an extraction blank and the five sample types following sample preparation.

### **Optional Solid Phase Extraction**

A dispersive SPE cleanup was used to achieve a 10-fold lower level of quantitation. Four replicate samples of the egg matrix were spiked with the PFAS analyte mix at the 0.1 ng/g level and processed by the QuEChERs procedure. Following extraction,  $500\,\mu\text{L}$  of the acetonitrile phase was diluted with  $15\,\text{mL}$  of water and loaded onto a preconditioned, weak-ion-exchange SPE tube (Phenomenex Strata®-X-AW 200 mg). The analytes of interest were then eluted with  $4\,\text{mL}$  of  $0.3\,\%$  NH $_4\text{OH-acetonitrile.The}$  eluate was evaporated to dryness, reconstituted with  $500\,\mu\text{L}$  of acetonitrile and transferred to an LC autosampler vial for analysis.

### **Optional Solid Phase Extraction**

A dispersive SPE cleanup was used to achieve a 10-fold lower level of quantitation. Four replicate samples of the egg matrix were spiked with the PFAS analyte mix at the 0.1 ng/g level and processed by the QuEChERs procedure. Following extraction,  $500\,\mu\text{L}$  of the acetonitrile phase was diluted with 15 mL of water and loaded onto a preconditioned, weak-ion-exchange SPE tube (Phenomenex Strata-X-AW 200 mg). The analytes of interest were then eluted with 4 mL of 0.3 % NH $_{\!_4}\text{OH-acetonitrile}$ . The eluate was evaporated to dryness, reconstituted with 500  $\mu\text{L}$  of acetonitrile and transferred to an LC autosampler vial for analysis.

### LC-MS/MS Analysis

The chromatography was performed on an Agilent® 1290 UH-PLC system. The LC column employed was a Phenomenex Luna™ Omega 1.6 µL PS C18 operating at 40 degrees Celsius with a flow rate of 0.55 mL/min and an injection volume of 20 µL. The mass spectrometer used was an Agilent 6460 QQQ. Various LC-MS/MS conditions were explored and an ammonium acetate/acetonitrile gradient (**Table 2**) proved to be optimum, resulting in a run time of approximately 4 minutes.

### **Results and Discussion**

System calibration showed a linear dynamic response from 0.05 ppb – 1000 ppb with a lower limit of quantization of 0.05 ppb as shown in **Figure 3** and a calibration chromatogram at the 0.05 ppb level is shown in **Figure 4**. Recovery data for the five matrix types is summarized in **Figures 5–9**. Four replicates of each matrix were spiked at the 1 ng/g level and prepared for analysis as described above (but were not subjected to the solid phase extraction process). **Figure 10** presents the recovery data for four replicates of the egg matrix spiked at 0.1 ng/g and prepared as described above, but with the addition of the solid phase extraction step to increase method sensitivity.

The recovery data show good recovery for all five matrices spiked at the 1 ng/g level, with most analytes falling into the 80% - 120% recovery range. Precision is generally somewhat poorer for the higher fat dairy products than for the lower fat matrices. The recoveries on tuna fish are particularly good, considering the complexity of the matrix. In comparing the analyte recoveries from eggs at the 1 ng/g and 0.1 ng/g levels (Figure 9 and Figure 10), both show comparable recoveries although, as expected, the higher spike level shows greater precision. Overall, the data suggest that the method has sufficient accuracy and precision to potentially be used to assess environmental PFAS contamination of food products. Clearly, this is preliminary data and further development and multi-laboratory validation would be required to demonstrate such a purpose. However, the data clearly show that current sample preparation techniques, coupled with the power of advanced chromatography and triple-quad mass spectrometry represent a suitable workflow.

#### The Sequel

The earlier discussion showed the use of current analytical technology to address the challenge of environmental PFAS contamination of the food supply. However, care should be taken since experience with analytical chemistry teaches us that we will inevitably be facing further analytical challenges from the realm of the "unknown-unknowns".

In PFAS analysis, we are currently discussing a target analyte list of 20, 30 or 40 compounds? However, the number of compounds in the PFAS universe has been estimated at 5000 - and even as high as 8,000 - which doesn't include potential degradation products. Toxicity is largely a function of the unique chemical and configurational state of a molecule that controls the biochemical interaction with the organism. So, there is much more analytical work to identify the most important PFAS compounds from a toxicity perspective.

Excellent work is being done with accurate mass and advanced data analysis to give us a broader understanding of the chemical complexity of the PFAS universe. However, given the complexity and extent of the problem of environmental PFAS contamination, it is clear that a lot of hard work has yet to be done.

#### **Acknowledgements**

The contribution of Dr. Agustin Pierri and his team at Weck Laboratories, City of Industry, California, USA is gratefully acknowledged.



Figure 1.
Pathway Model for Environmental Transmission of PFAS to Food and Consumer

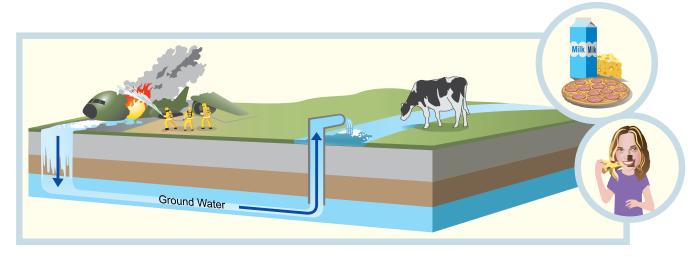
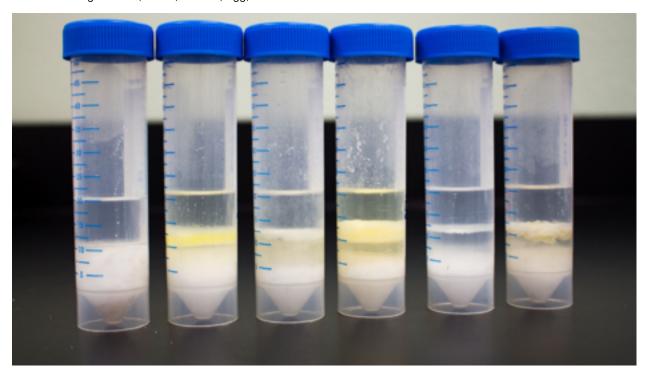


Figure 2.
Samples after QuEChERs Cleanup:
From Left to Right: Blank, Butter, Cheese, Egg, Milk and Fish



# **Applications**

Figure 3.
System Calibration Dynamic Range (0.05 – 1000 ppb)

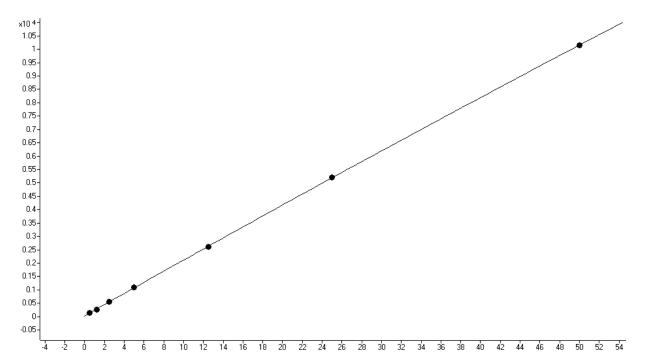


Figure 4.
Chromatogram of 0.05 ppb Lower Limit of Quantization Standard

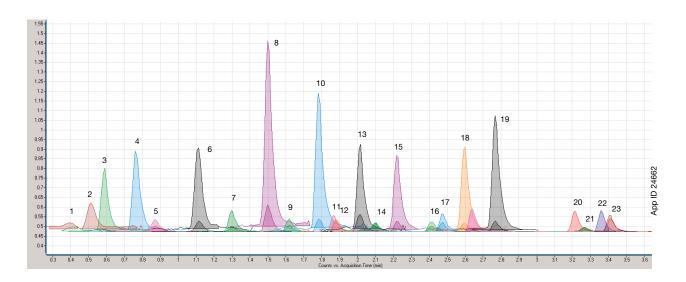


Figure 5.
Milk Recoveries (QuEChERs: 1 ng/g, n=4)

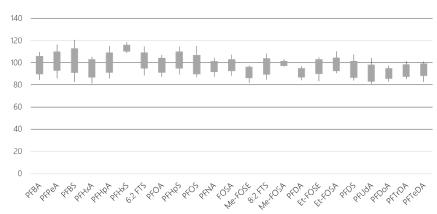


Figure 6.
Butter Recoveries (QuEChERs: 1 ng/g, n=4)

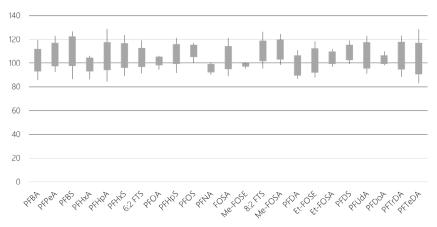


Figure 7.
Tuna Recoveries (QuEChERs: 1 ng/g, n=4)

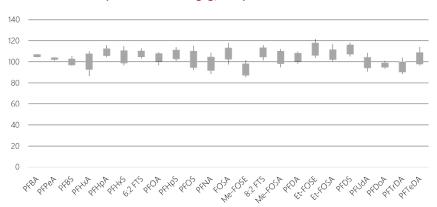


Figure 8.
Cheese Recoveries (QuEChERs:1 ng/g, n=4)

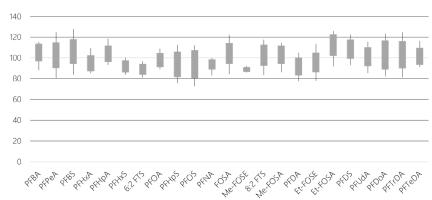


Figure 9. Egg Recoveries (QuEChERs: 1 ng/g, n=4)

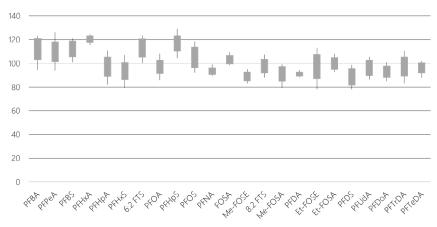
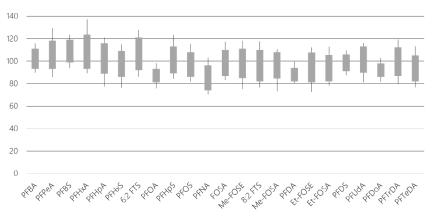


Figure 10. Egg Recoveries (QuEChERs + SPE: 0.1 ng/g, n=4)



## Table 1. PFAS Analyte List

Analytes:		
1. PFBA	9. PFHpS	17. Et-FOSE
2. PFPeA	10. PF0S	18. Et-FOSA
3. PFBS	11. PFNA	19. PFDS
4. PFHxA	12. F0SA	20. PFDS
5. PFHpA	13. Me-FOSE	21. PFDoA
6. PFHxS	14. 8:2 FTS	22. PFTrDA
7. 6:2 FTS	15. Me-FOSA	23. PFTeDA
8. PFOA	16. PFDA	

## **Table 2.** *LC-MS/MS Conditions*

Column: Luna™ Omega 1.6µm PS C18

**Dimensions:** 100 x 2.1 mm **Part No.:** 00D-4752-AN

Mobile Phase: A: 5 mM Ammonium Acetate in Water

B: Acetonitrile

Gradient: Time (min) % B
0 40
0.5 40
3 90

3 90 3.1 100 4 100 Flow Rate: 0.55 mL/min

Temperature: 40 °C
UHPLC System: Agilent® 1290
Detection: Agilent 6460 QQQ

Analytes: 1. PFBA 9. PFHpS 17. Et-FOSE
2. PFPeA 10. PFOS 18. Et-FOSA

PFBS
 PFNA
 PFDS
 PFHXA
 FOSA
 PFUdA
 PFHPA
 Me-FOSE
 PFDOA
 PFHXS
 S2 FTS
 Me-FOSA
 PFTDA
 Me-FOSA

8. PFOA 16. PFDA

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## 2. PFAS in Food Contact Materials

# Identification and Quantification of PFAS in Food Contact Materials using MRM<sup>HR</sup> Workflow on X500R QTOF System

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#### Introduction

In comparison to other surfactants, perfluorinated alkyl substances (PFAS) have stable physiochemical structures with hydrophobic and oleophobic properties. They are widely used in industrial and consumer products like plastic packaging materials for food and as coating in non-stick pans. Due to their chemical stability and low reactivity, PFAS are highly resistant to degradation even in living organisms and can therefore be accumulated in the food chain. Human exposure to PFAS residues has been implicated in incidences of cancer, obesity, endocrine system disruption and other adverse health effects. [1]

With the rapid growth in the food delivery industry in China (and globally) in the past two years, one-time-use plastic packaging materials are widely used by merchants due to their low cost and high durability [2]. One-time-use plastic has become a source of public concern and environmental pollution. Given the tremendous persistence of PFAS in the environment and the adverse effect on human health, monitoring of PFAS residue has gained traction in China and elsewhere.

In China, the level of PFOS and PFOA in food contact materials and products is regulated according to the latest National Food Safety Standard (GB 31604.35-2016). The detection limit is set at 1.0 ng/g while the quantification limit is set at 2.0 ng/g. In 2006, the European Union (EU) has set a regulation that the level of PFOS in finished products should not exceed 0.005 % of the product mass.

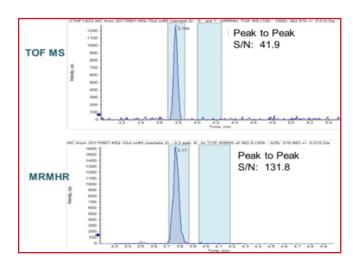


Figure 1.
Signal-to-Noise Comparison of PFHpA using TOF-MS and MRMHR Data Using a Post Spiked 0.2 ppb Matrix Blank

Monitoring the transition and the high resolution fragment ion results in greater specificity and reduced baseline, so signal-to-noise demonstrates marked improvement and method sensitivity is maximized.



The X500R QTOF system has the industry's fastest scanning speed, allowing for the implementation of the unique MRM<sup>HR</sup> acquisition mode to provide excellent quantitative performance using high-resolution MS/MS data. This approach to quantitation with LC-QTOF-MS/MS minimizes matrix interferences and the patented Turbo V<sup>™</sup> ion source with curtain gas interface, twin sprayer technology and built-in automatic calibration system help to improve and maintain instrument robustness and maintain high mass accuracy results. The high resolution MS/MS spectra can also be used for qualitative analysis by calculating the ion ratio for confirmation, thus reducing false positives by taking advantage of the data acquired on the LC-QTOF platform.

### **Key Workflow Advantages**

- PFAS quantitation using an easily established method and minimal method development
- 10-minute run time using a Phenomenex Kinetex® C18 column demonstrates separation of PFAS targets
- MRM<sup>HR</sup> workflow using MS/MS for selectivity vs high resolution TOF MS mode provides improved signal-to-noise
- QTOF technology can be utilized for quantitative analysis of PFAS suite without compromising method performance (excellent sensitivity, linearity demonstrated)

## 2. PFAS in Food Contact Materials (continued)

#### Methods

### **Sample Preparation**

The food packaging material to be tested is cut into small pieces. For coating sample, scrape it with a small knife. The sample preparation procedure was adapted from National Standard of China (document number GB 31604.35-2016) which is implemented on 19 April 2017 (**Figure 2**).

A total of eight samples were collected as test samples which include disposable meal box, plastic bag, beverage bottle, coating of non-stick pan, etc. Packaging materials in the collected samples were mainly polyethylene, polystyrene and polytetra-fluoroethylene.

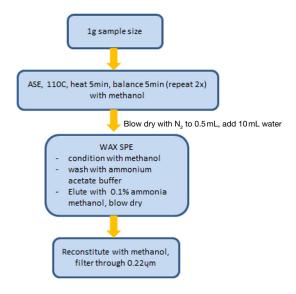


Figure 2. Extraction and Clean-up Process Flow Diagram

### Chromatography

Using the SCIEX® ExionLC $^{\infty}$  AD System with a Phenomenex Kinetex®, 2.6 µm C18, 100 X 2.0 mm, compounds were separated using a gradient elution with mobile phase A of 5 mM NH $_4$ AC in water and mobile phase B of 5 mM NH $_4$ AC in methanol (flow rate of 0.3 mL/min, column temperature 40  $^{\circ}$ C).

### **Mass Spectrometry**

The SCIEX X500R QTOF System was used to analyse the compounds operating in negative ion polarity using the Scheduled MRM<sup>HR</sup> acquisition mode (**Table 1**). Source conditions were as follows: CUR of 30psi; CAD of 7; IS of -4500V; Temp 500 °C; GS1 of 50psi; GS2 of 55psi.

### **Data Processing**

All data was processed with SCIEX OS Software.

## Table 1. Scheduled MRM<sup>HR</sup> Method Setup in SCIEX OS

Unique RTs can be defined for each transition for each analyte.

	Compound	Group	Precursor	Fragment	Accumulation	Declustering	Collision	Retention
	ID	name	ion (Da)	ion (Da)	time (sec)	potential (V)	energy (V)	time (min)
1	PFBA	PFBA	212.90	168.9000	0.0600	-80	-35	2.55
2	PFPeA 1	PFPeA	262.90	218.9000	0.0600	-80	-35	3.22
3	PFPeA 2	PFPeA	262.90	69.0000	0.0600	-80	-35	3.22
4	PFBS 1	PFBS	298.90	80.0000	0.0600	-80	-35	3.29
5	PFBS 2	PFBS	298.90	99.0000	0.0600	-80	-35	3.29
6	PFHxA 1	PFHxA	312.90	268.9000	0.0600	-80	-35	3.55
7	PFHxA 2	PFHxA	312.90	119.0000	0.0600	-80	-35	3.55
8	PFHxS 1	PFHxS	362.90	318.9000	0.0600	-80	-35	3.79
9	PFHxS 2	PFHxS	362.90	168,9000	0.0600	-80	-35	3.79

### Establishing the Scheduled MRMHR Quantitative Method

The SCIEX OS software is fully automated with a user-friendly interface, greatly reducing the time to establish the acquisition method. The MRM parameters can be set up easily in two different ways. For compounds which are in MS/MS spectral library, fragment ions can be imported easily from the library to build the MRMHR method list. Up to 5 fragment ions can be imported at the same time using a single click. For compounds not found in the spectral library, spectra can be added easily to the library using TOF MS-IDA-MS/MS data acquired for standards of the desired targets.

MRM parameters like retention time, declustering potential (DP) and collision energy (CE) from an existing triple quadrupole method are fully transferrable.

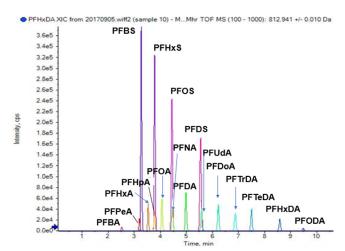


Figure 3.
TOF MS Extracted Ion Chromatogram of 17 PFAS
Good separation was achieved for most of the 17 PFAS compounds analysed.

## 2. PFAS in Food Contact Materials (continued)

### MRM<sup>HR</sup> Quantitation of PFAS

Chromatogram of 17 PFAS utilizing extracted precursor ion data from TOF-MS scan are shown (**Figure 3**).

### **High Selectivity Data**

Comparing 0.2 ppb post spiked in matrix blank, PFHpA show higher selectivity in MRM<sup>HR</sup> mode as compared to TOF-MS mode for quantification (**Figure 1**). Monitoring the high resolution fragment ion from the full scan MS/MS data collected provides greater specificity and reduced baseline, so signal-to-noise demonstrates marked improvement and method sensitivity is maximized.

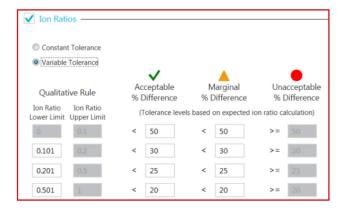
### **Linearity and Accuracy**

The 17 monitored PFAS demonstrate good linearity and accuracy (**Figure 4**) with the correlation coefficients above 0.99. Accuracy values are within the permissible deviation range for LOD and LOQ according to the national standards.

### **Ion Ratio Calculations**

Ion ratios can be easily calculated using the SCIEX® OS software. Ion ratio confirmation can be visually displayed in the chromatogram and result table. Depending on the requirement, the confirmation tolerance can be defined using either constant tolerance or variable tolerance as shown in **Figure 5**.





### Figure 5.

### **Setting up Tolerance for Ion Ratios Confirmation**

Constant tolerance (same percent difference from measured standard ion ratio) or variable tolerance (varying percent difference dependant on concentration level) can be utilized when determining whether an unknown same meets the criteria for qualitative analyte identification by ion ratio confirmation. Different levels of percent difference can be defined by the user to be flagged as within "Acceptable," "Marginal," or "Unacceptable."

### **Detection of PFAS in Food Contact Materials**

SCIEX OS software combines both qualitative and quantitative results in one single interface (**Figure 6**). The result table show the retention time, concentration, peak area, ion ratio confirmation and the mass error of 0.9 ppm for a sample tested positive with PFOA.

Among the eight samples, eight types of PFAS were detected as shown in **Table 2**. Two out of eight samples have levels which exceeded regulated level of 1 ng/g by national standard. Most of the detected PFAS are the acid derivatives of PFOA and primarily found in non-stick pan coating and disposable meal boxes. The number of actual samples collected in this test is rather small; hence statistically it does not imply that all related products are unsafe for consumers.

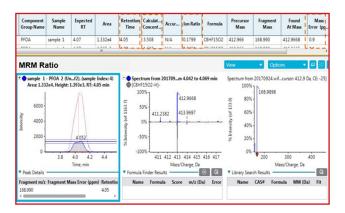


Figure 6. PFOA Results in Actual Sample

## 2. PFAS in Food Contact Materials (continued)

#### Summary

The SCIEX® X500R QTOF system and SCIEX OS software brings powerful performance capabilities for routine testing of PFAS. The unique MRMHR quantification method enables high selectivity even in real sample with matrix interference. This improves the detection and quantification of PFAS which can meet the EU regulation and national standards in China.

Although the concentration of PFAS in most of the test samples falls below the regulated level, the detection rate of perfluorinated alkyl substances is relatively high indicating that the quality of food contact/packaging materials may pose potential risks to consumer's health.

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- GB 31604.35-2016, National Food Safety Standard Food contact materials and products Determination of perfluorooctane sulfonate (PFOS) and perfluorooctanoic acid (PFOA).

Table 2.
PFAS Content in Different Food Contact Samples

### Detected Amount (ng/g)

					· ·			
	PFHxA	PFHpA	PFOA	PFDA	PFuDA	PFDoA	PFTrDA	PFTeDA
Meal box 1	0.14	0.16	3.15	-	-	-	-	-
Meal box 2	-	-	3.12	-	-	-	-	-
Plastic bag 1	-	-	-	-	-	-	-	-
Plastic bag 2	-	-	-	-	-	-	-	-
Drink bottle 1	-	-	-	-	-	-	-	-
Drink bottle 2	-	-	-	-	-	-	-	-
Non-stick pan 1	-	-	-	0.11	0.15	0.13	0.15	-
Non-stick pan 2	-	-	-	-	-	-	-	0.17

<sup>-</sup> Falls below the detection level of this method.

Figure 4.
Calibration Curve of 17 PFAS with Acceptable Accuracy and Linear Response

Sample Name	Sample Type	Compo Name	Component Group Na	Actual Conce	Expected RT	Area	Retention Time	Used	Calculated Concentration	Accuracy
0.1	Standard	PFDS	PFDS	0.10	5.55	6.243e2	5.54	<b>V</b>	0.101	101.37
0.2	Standard	PFDS	PFDS	0.20	5.55	1.121e3	5.55	V	0.199	99.64
).5	Standard	PFDS	PFDS	0.50	5.55	2.634e3	5.54	V	0.497	99.46
	Standard	PFDS	PFDS	1.00	5.55	5.120e3	5.54	V	0.987	98.71
2	Standard	PFDS	PFDS	2.00	5.55	1.041e4	5.52	V	2.028	101.42
;	Standard	PFDS	PFDS	5.00	5.55	2.570e4	5.53	V	5.040	100.81
10	Standard	PFDS	PFDS	10.00	5.55	4.971e4	5.53	V	9.772	97.72
20	Standard	PFDS	PFDS	20.00	5.55	1.025e5	5.53	V	20.174	100.87
Calibration for Calibration fo	PFPEA 2: Not end PFBS: y = 2236.25 PFBS 2: y = 1383 PFHxA: y = 11330 PFHxA 2: y = 467	ough data p 5077 x + -1 .61894 x + 5.01544 x + 7.58753 x +	oints. 6.05946 (r = 0 135.80528 (r -523.72664 (r 140.21164 (r	.99982, r <sup>2</sup> = 0.99850, r = 0.9994 = 0.99694	= 0.99963) ( r <sup>2</sup> = 0.99699 5, r <sup>2</sup> = 0.9988 , r <sup>2</sup> = 0.99389	(weighting: 1 / x) () (weighting: 1 / () (weighting: 1 / () (weighting: 1	/x) L/x) /x)	0.0	5-20p	pb
Calibration for Calibration fo	PFPA 2: Not end PFBS: y = 2236.2: PFBS 2: y = 1383 PFHXA 2: y = 13133 PFHXA 2: y = 467 PFHPA: y = 1540 PFHPA 2: y = 180 PFHPA 2: y = 2465. PFHXS 2: y = 566 PFOA: y = 15044 PFOA 2: y = 191	ough data p 5077 x + -1 .61894 x + 5.01544 x + 5.015461 x + 0.11861 x + 0.8.78206 x 64427 x + 1 .74618 x + .28158 x + 1.93693 x +	oints. 6.05946 (r = 0 135.80528 (r) -523.72664 ( 140.21164 (r) -368.46808 ( +222.24516 ( 57.59648 (r) 42.25411 (r) -677.06515 (r) 205.51738 (r)	.99982, r <sup>2</sup> = 0.99850, r = 0.9994 = 0.99694 (r = 0.9986 0.99943, 0.99740, r = 0.99830 = 0.99956	= 0.99963) ( r <sup>2</sup> = 0.99699 5, r <sup>2</sup> = 0.9988 8, r <sup>2</sup> = 0.9988 8, r <sup>2</sup> = 0.9988 5, r <sup>2</sup> = 0.9988 2 = 0.99886 2 = 0.99886 3 = 0.99886 4 = 0.99886 6 = 0.999886 6 = 0.999886 7 = 0.999886	weighting: 1 / x) ) (weighting: 1 / x) 89) (weighting: 1 90) (weighting: 1 96) (weighting: 1 97) (weighting: 1 / (yeighting: 1 / 1) (weighting: 1 98) (weighting: 1 99) (weighting: 1 99) (weighting: 1 90) (weighting: 1 90) (weighting: 1 90) (weighting: 1	/x) //x) /x) 1/x) 1/x) 1/x) x) x) /x)	).0 R <sup>2</sup>	5-20p 2 = 0.9	pb 9
Calibration for for	PFPA 2: Not end PFBS: y = 2236.2: PFBS 2: y = 1383 PFHXA 2: y = 13133 PFHXA 2: y = 467 PFHPA: y = 1540 PFHPA 2: y = 180 PFHPA 2: y = 2465. PFHXS 2: y = 566 PFOA: y = 15044 PFOA 2: y = 191	ough data p 5077 x + -1 .61894 x + 5.01544 x + 5.015461 x + 0.11861 x + 0.8.78206 x 64427 x + 1 .74618 x + .28158 x + 1.93693 x +	oints. 6.05946 (r = 0 135.80528 (r) -523.72664 ( 140.21164 (r) -368.46808 ( +222.24516 ( 57.59648 (r) 42.25411 (r) -677.06515 (r) 205.51738 (r)	.99982, r <sup>2</sup> = 0.99850, r = 0.9994 = 0.99694 (r = 0.9986 0.99943, 0.99740, r = 0.99830 = 0.99956	= 0.99963) ( r <sup>2</sup> = 0.99699 5, r <sup>2</sup> = 0.9988 8, r <sup>2</sup> = 0.9988 8, r <sup>2</sup> = 0.9988 5, r <sup>2</sup> = 0.9988 2 = 0.99886 2 = 0.99886 3 = 0.99886 4 = 0.99886 6 = 0.999886 6 = 0.999886 7 = 0.999886	weighting: 1 / x) ) (weighting: 1 / x) 89) (weighting: 1 90) (weighting: 1 96) (weighting: 1 97) (weighting: 1 / (yeighting: 1 / 1) (weighting: 1 98) (weighting: 1 99) (weighting: 1 99) (weighting: 1 90) (weighting: 1 90) (weighting: 1 90) (weighting: 1	/x) //x) /x) 1/x) 1/x) 1/x) x) x) /x)	).0 R <sup>2</sup>	5-20pp	pb 9
Calibration for Calibration fo	PFPA 2: Not end PFBS: y = 2236.2: PFBS 2: y = 1383 PFHXA 2: y = 13133 PFHXA 2: y = 467 PFHPA: y = 1540 PFHPA 2: y = 180 PFHPA 2: y = 2465. PFHXS 2: y = 566 PFOA: y = 15044 PFOA 2: y = 191	ough data p 5077 x + -1 .61894 x + 5.01544 x + 5.015461 x + 0.11861 x + 0.8.78206 x 64427 x + 1 .74618 x + .28158 x + 1.93693 x +	oints. 6.05946 (r = 0 135.80528 (r) -523.72664 ( 140.21164 (r) -368.46808 ( +222.24516 ( 57.59648 (r) 42.25411 (r) -677.06515 (r) 205.51738 (r)	.99982, r <sup>2</sup> = 0.99850, r = 0.9994 = 0.99694 (r = 0.9986 0.99943, 0.99740, r = 0.99830 = 0.99956	= 0.99963) ( r <sup>2</sup> = 0.99699 5, r <sup>2</sup> = 0.9988 8, r <sup>2</sup> = 0.9988 8, r <sup>2</sup> = 0.9988 5, r <sup>2</sup> = 0.9988 2 = 0.99886 2 = 0.99886 3 = 0.99886 4 = 0.99886 6 = 0.999886 6 = 0.999886 7 = 0.999886	weighting: 1 / x) ) (weighting: 1 / x) 89) (weighting: 1 90) (weighting: 1 96) (weighting: 1 97) (weighting: 1 / (yeighting: 1 / 1) (weighting: 1 98) (weighting: 1 99) (weighting: 1 99) (weighting: 1 90) (weighting: 1 90) (weighting: 1 90) (weighting: 1	/x) //x) /x) 1/x) 1/x) 1/x) x) x) /x)	).0 R <sup>2</sup>	5-20p	pb 9
Calibration for for for Calibration for	PFPA 2: Not end PFBS: y = 2236.2: PFBS 2: y = 1383 PFHXA 2: y = 13133 PFHXA 2: y = 467 PFHPA: y = 1540 PFHPA 2: y = 180 PFHPA 2: y = 2465. PFHXS 2: y = 566 PFOA: y = 15044 PFOA 2: y = 191	ough data p 5077 x + -1 .61894 x + 5.01544 x + 5.015461 x + 0.11861 x + 0.8.78206 x 64427 x + 1 .74618 x + .28158 x + 1.93693 x +	oints. 6.05946 (r = 0 135.80528 (r) -523.72664 ( 140.21164 (r) -368.46808 ( +222.24516 ( 57.59648 (r) 42.25411 (r) -677.06515 (r) 205.51738 (r)	.99982, r <sup>2</sup> = 0.99850, r = 0.9994 = 0.99694 (r = 0.9986 0.99943, 0.99740, r = 0.99830 = 0.99956	= 0.99963) ( r <sup>2</sup> = 0.99699 5, r <sup>2</sup> = 0.9988 8, r <sup>2</sup> = 0.9988 8, r <sup>2</sup> = 0.9988 5, r <sup>2</sup> = 0.9988 2 = 0.99886 2 = 0.99886 3 = 0.99886 4 = 0.99886 6 = 0.999886 6 = 0.999886 7 = 0.999886	weighting: 1 / x) ) (weighting: 1 / x) 89) (weighting: 1 90) (weighting: 1 96) (weighting: 1 97) (weighting: 1 / (yeighting: 1 / 1) (weighting: 1 98) (weighting: 1 99) (weighting: 1 99) (weighting: 1 90) (weighting: 1 90) (weighting: 1 90) (weighting: 1	/x) //x) /x) 1/x) 1/x) 1/x) x) x) /x)	0.0 R <sup>2</sup>	5-20pp = 0.9	pb 9

# Product Guide for PFAS Analysis

### Phenomenex PFAS Products Referenced or Applicable in Official Methods

Regulatory Method	Product	Part Number
	PFAS CRM EPA 537.1 mix 1mL 2µg/mL in methanol	<u>AL0-101839</u>
USEPA 537.1: Determination of Selected Per-and Polyfluorinated Alkyl Substances in Drinking Water by	PFAS CRM EPA 533 + 537.1 mix 1mL 2µg/mL in methanol	<u>AL0-101840</u>
Solid Phase Extraction and Liquid Chromatography/ Tandem Mass Spectrometry (LC/MS/MS) (5)	Strata™ SDB-L 500 mg/6 mL	8B-S014-HCH
	Gemini™ 3 µm C18, 50 x 3 mm or	00B-4439-B0
	Luna™ Omega 1.6 µm PS C18 100 x 2.1 mm	00D-4752-AN
	PFAS CRM EPA 533 mix 1mL 2µg/mL in methanol	AL0-101838
USEPA Method 533: Determination of Per- and Polyfluoroalkyl Substances in Drinking Water by Isotope	PFAS CRM EPA 533 + 537.1 mix 1mL 2µg/mL in methanol	<u>AL0-101840</u>
Dilution, Anion Exchange Solid Phase Extraction and LC-MS/MS. (1)	Strata-X-AW 500 mg/6 mL	8B-S038-HCH
	Gemini 3 µm C18 50 x 2 mm or	<u>00B-4758-Y0</u>
	Luna Omega 1.6 µm PS C18 100 x 2.1 mm	00D-4752-AN
US Food and Drug Administration: Determination of 16 Perfluoroalkyl and Polyfluoroalkyl Substances(PFAS) in Food using Liquid Chromatography-Tandem Mass Spectrometry (LC-MS/MS). (2)	Strata-XL-AW 200 mg/3 mL	8B-S051-FBJ
US Department of Agriculture: Screening, Determiation and Confirmation of PFAS by UPLC-MS-MS (3)	Luna C8(2) 3 µm 50 x 2 mm	00B-4248-B0
US Department of Defense: Quality Systems Manual (QSM) for Environmental Laboratories (4)	<b>Strata PFAS (WAX/GCB)</b> 200 mg/50 mg/6 mL, 30/box 500 mg/50 mg/6 mL, 30/box	CSO-9207 CSO-9208
References	Gemini 3 µm C18 50 x 2 mm	<u>00B-4439-B0</u>

- Method 537.1: Determination of Selected Per- and Polyfluorinated Alkyl Substances in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS) | Science Inventory
- 2. Method 533: Determination of Per- and Polyfluoroalkyl Substances in <u>Drinking Water by Isotope Dilution Anion Exchange Solid Phase Extraction</u> and Liquid Chromatography/Tandem Mass Spectrometry | Methods Approved to Analyze Drinking Water Samples to Ensure Compliance with Regulations | US EPA
- 3. Determination of 16 Perfluoroalkyl and Polyfluoroalkyl Substances in Food using Liquid Chromatography-Tandem Mass Spectrometry (fda.gov)
- Screening, Determination and Confirmation of PFAS by UPLC-MS-MS
- 5. https://denix.osd.mil/edqw/documents/manuals/qsm-version-5-3-final/

### **Recommended HPLC Products for Routine PFAS Analysis**

Description and Function	Product	Part Number
	Kinetex™ 5 µm EV0 C18 100 x 2.1 mm	00D-4633-AN
Analytical Column (UHPLC)	Luna Omega C18 1.6 µm 50 x 2.1	00B-4752-AN
	Gemini 3 µm C18 50 x 3 mm	00B-4439-Y0
Analytical Column	Gemini 3 µm C18 50 x 3 mm	00B-4439-Y0
Analytical Column (> 100 µL injection)	Gemini 3 µm C18 100 x 3 mm	00D-4439-Y0
Analytical Column (improved. Imwt acids)	Luna Omega 3 µm PS C18 50 x 3 mm	<u>00B-4758-Y0</u>
Delay Column	Kinetex 5 µm EVO C18, 50 x 2.1 mm	00B-4633-AN 00A-4252-Y0
SecurityGuard	Luna Omega PS C18 4 x 3.0/10 pack for ID: 3.2-8.0 mm 4 x 2.0/10 pack for ID: 2.0-3.0 mm	AJ0-7606 AJ0-7605

# Product Guide for PFAS Analysis (continued)

### **Recommended SPE Products**

Description and Function	Product	Part Number
SPE Cartridge for EPA 537.1	Strata™ SDB-L 500 mg/6 mL, 30/box	8B-S014-HCH
SPE Cartridge for EPA 533	Strata-X-AW 33um Polymeric Weak Anion, 500 mg/6 mL, 30/box	8B-S038-HCH
SPE Cartridge (Rev. Phase, High Perf.)	Strata-XL 500 mg/6 mL, 30/box	<u>8B-S043-HCH</u>
SPE Stacked Cartridge (DOD QSM 5.3)	Strata PFAS (WAX/GCB) 200 mg/50 mg/6 mL, 30/box	<u>CS0-9207</u>
SPE Stacked Cartridge (DOD QSM 5.3)	Strata PFAS (WAX/GCB) 500 mg/50 mg/6 mL, 30/box	CS0-9208
SPE Cartridge (WAX for DOD QSM 5.3)	Strata-XL-AW 500 mg/6 mL, 30/box	8B-S051-HCH
GCB** Cartridge (GCB for DOD QSM 5.3)	Strata GCB 250 mg/6 mL, 30/box	<u>8B-S528-FCH</u>
SPE Cartridge (WAX* for FDA Method)	Strata-XL-AW 100 µm 200 mg/3 mL, 50/box	<u>8B-S051-FBJ</u>
(*WAX = Weak Anion Exchange) (**GCB = Graphitized Carbon Black)		

### **Recommended QuEChERs Products**

Description and Function	Product	Part Number
QuEChERs Extraction (Soil/Sediment)	roQ QuEChERs Extraction Kit	KS0-8911
QuEChERs dSPE (Soil/Sediment)	roQ QuEChERs dSPE Kit, 15 mL	KS0-9516
QuEChERs Extraction (Dairy/Eggs/Fish)	roQ QuEChERs Extraction Kit	KS0-8910
QuEChERs dSPE (Dairy/Eggs/Fish)	roQ QuEChERs dSPE Kit	KS0-9511

### **Recommended Accessories**

Description and Function	Product	Part Number
SPE Sample Reservoir	75 mL Sample Reservoir	<u>H0-7005</u>
Large Volume SPE	Adaptor Cap for 12,20, 60 mL SPE Tubes	<u>AH0-7379</u>
Autosampler Vials	Polypropylene, 300 µm + PE Starburst Cap	AR0-9995-12-C
Polypropylene Vials	Vial 9mm Screw Thd PP 2mL, 1000 Pk	AR0-89C7-13
Vial Caps	Cap 9 mm Solid Top Black Unlined	8B-S528-FCH
PEEK Capillary Tubing	Capillary Tubing Kit, Various Sizes	AT0-1964
PEEK Tubing Cutter	Cutter for PEEK Capillary Tubing	AT0-1110

# Strata<sup>™</sup> Solid Phase Extraction (SPE)

### Strata-X

**Ordering Information** 

Format	Sorbent Mass	Part Number	Unit
Tube			
@'strata ==	30 mg	8B-S100-TAK**	1 mL (100/box)
	30 mg	8B-S100-TBJ	3 mL (50/box)
	60 mg	8B-S100-UBJ**	3 mL (50/box)
	100 mg	8B-S100-EBJ	3 mL (50/box)
	100 mg	8B-S100-ECH	6 mL (30/box)
	200 mg	8B-S100-FBJ	3 mL (50/box)
	200 mg	8B-S100-FCH	6 mL (30/box)
	500 mg	8B-S100-HBJ	3 mL (50/box)
	500 mg	8B-S100-HCH	6 mL (30/box)
Giga™ Tube			
⊜strata	500 mg	8B-S100-HDG	12 mL (20/box)
Sur planting at	1 g	8B-S100-JDG	12 mL (20/box)
	1 g	8B-S100-JEG	20 mL (20/box)
	2 g	8B-S100-KEG	20 mL (20/box)
	5 g	8B-S100-LFF	60 mL (16/box)
Teflon® Tube			
⊜strata =	200 mg	8B-S100-FBJ-T	3 mL (50/box)
- corporated	200 mg	8B-S100-FDG-T	12 mL (20/box)

### **On-line Extraction Cartridge**

Description	Part Number	Unit/Box	
Strata-X on-line extraction cartridge, 20 x 2.0 mm	00M-S033-B0-CB	ea	
Cartridge holder, 20 mm	<u>CH0-5845</u>	ea	

<sup>\*\*</sup>Tab-less tubes available. Contact Phenomenex for details

### Strata-XL

**Ordering Information** 

Format	Sorbent Mass	Part Number	Unit
Tube			
Satrata III.	30 mg	8B-S043-TAK	1 mL (100/box)
	60 mg	8B-S043-UBJ	3 mL (50/box)
	100 mg	8B-S043-EBJ	3 mL (50/box)
	200 mg	8B-S043-FBJ	3 mL (50/box)
	200 mg	8B-S043-FCH	6 mL (30/box)
	500 mg	8B-S043-HCH	6 mL (30/box)
Giga Tube			
⊜strata	2 g	8B-S043-KDG	12 mL (20/box)
Sur pharman	2 g	8B-S043-KEG	20 mL (20/box)
	5 g	8B-S043-LEG	20 mL (20/box)
	5 g	8B-S043-LFF	60 mL (16/box)
	10 g	8B-S043-MFF	60 mL (16/box)
	30 mg	8E-S043-TGB	2 Plates/Box

<sup>\*</sup> To control flow rate with Strata-XL, use a stopcock (AH0-6048) when processing samples with a vacuum manifold.

# Gemini™ pH Flexible LC Columns

### **Ordering Information**

3µm Microb	3µm Microbore, Minibore and MidBore™ Columns (mm)							SecurityGuard <sup>®</sup>	Cartridges (mm)	
Phases	50 x 1.0	20 x 2.0	30 x 2.0	50 x 2.0	100 x 2.0	150 x 2.0	50 x 3.0	100 x 3.0	150 x 3.0	4 x 2.0* /10pk
C18	00B-4439-A0	00M-4439-B0	<u>00A-4439-B0</u>	00B-4439-B0	00D-4439-B0	00F-4439-B0	00B-4439-Y0	00D-4439-Y0	00F-4439-Y0	<u>AJ0-7596</u>

for ID: 2.0-3.0 mm

3µm Analy	tical Columns (mm)		SecurityGuard™ Cartridges (mm)			
Phases	30 x 4.6	50 x 4.6	100 x 4.6	150 x 4.6	250 x 4.6	4 x 3.0* /10pk
C18	00A-4439-E0	00B-4439-E0	00D-4439-E0	00F-4439-E0	00G-4439-E0	AJ0-7597
						for ID: 3.2-8.0 mm



# Kinetex™ Core-Shell LC Columns

### **Ordering Information**

2.6 µm Micro LC Columns (mm)								
Phases	30 x 0.3	50 x 0.3	100 x 0.3	150 x 0.3	50 x 0.5	150 x 0.5		
EVO C18	_	00B-4725-AC	_	00F-4725-AC	00B-4725-AF	_		

2.6 µm MercuryMS™ LC-MS Cartridges (mm)							
Phases	20 x 2.0	20 x 4.0					
Biphenyl	00M-4622-B0-CE	00M-4622-D0-CE					



2.6 µm Minibor	e Columns (mm)					SecurityGuard ULTRA Cartridges <sup>‡</sup>
Phases	30 x 2.1	50 x 2.1	75 x 2.1	100 x 2.1	150 x 2.1	3/pk
EV0 C18	00A-4725-AN	00B-4725-AN	_	00D-4725-AN	00F-4725-AN	<u>AJ0-9298</u>
						for 2.1 mm ID

2.6 µm MidBore	2.6 µm MidBore™ Columns (mm)								
Phases	30 x 3.0	50 x 3.0	75 x 3.0	100 x 3.0	150 x 3.0	3/pk			
EVO C18	00A-4725-Y0	00B-4725-Y0	_	00D-4725-Y0	00F-4725-Y0	AJ0-9297			
						for 3.0 mm ID			





### **Ordering Information**

5 µm Minibor	e Columns (mm)				SecurityGuard™ ULTRA Cartridges‡
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk
EVO C18	00A-4633-AN	00B-4633-AN	00D-4633-AN	00F-4633-AN	<u>AJ0-9298</u>
					for 2.1 mm ID

5µm MidBore	™ Columns (mm)				SecurityGuard ULTRA Cartridges‡
Phases	30 x 3.0	50 x 3.0	100 x 3.0	150 x 3.0	3/pk
EV0 C18	00A-4633-Y0	00B-4633-Y0	00D-4633-Y0	00F-4633-Y0	<u>AJ0-9297</u>
					for 3.0 mm ID

5μm Analytic	al Columns (mm)				SecurityGuard ULTRA Cartridges‡
Phases	50 x 4.6	100 x 4.6	150 x 4.6	250 x 4.6	3/pk
EVO C18	00B-4633-E0	00D-4633-E0	00F-4633-E0	00G-4633-E0	<u>AJ0-9296</u>
					for 4.6 mm ID

\*SecurityGuard ULTRA Cartridges require holder, Part No.: AJO-9000 \*\*\*SemiPrep SecurityGuard Cartridges require holder, Part No.: AJO-9281 \*PREP SecurityGuard Cartridges require holder, Part No.: AJO-8223



# Luna™ One of The World's Leading LC Columns



### Luna C18

### **Ordering Information**

5µm MidBore and Analytical Columns (mm)							SecurityGuard™ C	artridges (mm)	
Phases	30 x 3.0	50 x 3.0	150 x 3.0	250 x 3.0	30 x 4.6	50 x 4.6	75 x 4.6	4 x 2.0*	4 x 3.0*
								/10pk	/10pk
C18(2)	00A-4252-Y0	00B-4252-Y0	00F-4252-Y0	00G-4252-Y0	00A-4252-E0	00B-4252-E0	00C-4252-E0	AJ0-4286	AJ0-4287
								for ID: 2.0-3.0 mm	3.2-8.0 mm

5μm Analytical and Semi-Prep Columns (mm)					SecurityGuard™ Cartridges (mm)		
Phases	100 x 4.6	150 x 4.6	250 x 4.6	250 x 10	4 x 3.0*	10 x 10 <sup>‡</sup>	
					/10pk	/3pk	
C18(2)	00D-4252-E0	00F-4252-E0	00G-4252-E0	00G-4252-N0	AJ0-4287	AJ0-7221	
					for ID: 3 2-8 0 mm	9-16 mm	

\*SecurityGuard™ Analytical Cartridges require holder, Part No.: <u>KJ0-4282</u> \*SemiPrep SecurityGuard™ Cartridges require holder, Part No.: <u>AJ0-9281</u>

### Luna Omega PS C18 and Luna C18

### **Ordering Information**

1.6 µm Microbore Columns (mm)							
Phases	50 x 1.0	100 x 1.0	150 x 1.0				
PS C18	_	00D-4752-A0	_				
C18	00B-4742-A0	00D-4742-A0	00F-4742-A0				

1.6 µm Minibore Columns (mm)				SecurityGuard™ ULTRA Cartridges <sup>‡</sup>	
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk
PS C18	00A-4752-AN	00B-4752-AN	00D-4752-AN	00F-4752-AN	AJ0-9508
C18	00A-4742-AN	00B-4742-AN	00D-4742-AN	00F-4742-AN	AJ0-9502
					for 2.1 mm ID

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