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Perfluoroalkyl Substances (PFAS) Analysis in Drinking Water, Sediments, and Food Samples by QuEChERS, SPE, and LC-MS/MS

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Introduction

Perfluoroalkyl substances (PFAS) are a class of highly stable synthetic organic compounds used in a wide variety of industrial and commercial applications including surface treatment for textiles, packaging materials, non-stick cookware, and fire fighting foams. PFAS are characterized by a hydrophobic fully fluorinated alkyl chain and a hydrophilic functional group. They are persistent in the environment due to the exceptional stability of the C-F bond. These have been detected throughout the global environment, food products, and even human plasma. PFAS are associated with various adverse health effects, they are bio-accumulative, ubiquitous, and their analysis level requirements are very low to account for an expected lifetime of exposure. There are several methods available for the extraction and analysis of PFAS in aqueous samples. However, very few procedures are available for extracting these compounds in solid matrices such as sediments and food samples.

Presented here are three methods making use of various sample preparation techniques for the analysis of PFAS. The methods include direct inject technique for dinking water, QuEChERS for sediment samples, and QuEChERS followed by SPE for food samples (milk, eggs, and fish tissue). All are validated procedures and make use of LC-MS/MS.

Water Samples





Figure 1. Representative Chromatogram of PFAS in Water Samples.



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The above application presents a method for the rapid analysis and quantitation of PFAS in water samples by UHPLC-MS/MS using the Luna Omega 1.6 µm PS C18 column. The high resolution and unique mixed-mode selectivity of the Luna Omega 1.6 µm PS C18 column results in excellent chromatography and peak shapes for PFAS in a very short four-minute run with solid retention and peak shape for PFBA and resolution of branched isomers

Sediment Samples

QuEChERS Extraction Protocol



Place an aliquot of the extract in an HPLC vial and dilute 1:1 with deionized water. The sample is now ready for analysis.

HPLC-MS/MS Conditions

Column: Dimensions: Part No.: Inline Filter: Delay Column: Dimensions: Part No.:	Gemini™ 3 µm C18 100 x 3 mm 00D-4439-Y0 KrudKatcher™ Ultra Luna™ 5 µm C18(2) 30 x 2.0 mm 00A-4252-B0		Analytes:	 PFBA PFPeA PFBS PFHxA PFPS PFHxS PFHxA PFHxA 8. PFHpA 8. PFHpS 	11 12 14 14 14 16 17 18
Mobile Phase:	A: 20 mM Ammoniun	n Acetate in Water		9. PFOA	1
	B: Methanol			10. PFOS	
Gradient:	Time (min) 0.00 1.50 8.00 8.10 12.0 12.5	% B 10 65 95 99 99 99 10	1		
Flow Rate:	0.6 mL/min				
Injection:	90 µL		2		
Temperature:	40 °C		4		J
Detector:	SCIEX [®] 5500 QTRA	D ®			
Detector:	MS/MS ESI Negative	∌ (sMRM)	1		

Figure 2. Extracted Ion Chromatogram of Sediments Spiked with 1.0 ng/g of the Target Analytes.



The above sediment extraction makes use of a modified QuEChERS method and is a fast, effective, and efficient way of extracting 19 PFAS from marine and river sediment matrices. The procedure significantly minimizes sample preparation time, solvent consumption, and overall cost of analysis. Executing a QuEChERS-like extraction and exploiting available kitted QuEChERS salts and dSPE products, effective, and accurate extraction of PFAS from solid and oily sediments enabled analysis down to low ng/g range.

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1. PFNA 2. PFOSA 3. PFNS 4 PFDA 5. PFDS 6. PFUdA . PFDoA 8. PFTrDA 9 PFTeDA





Food Samples (Dairy and Fish)

Sample	Preparation		HPLC-MS/M	S Conditions	
Step		Description	Column:	Luna Omega	
roQ Extraction Kit (Part No.: KS0-8910)			Dimensions: Part No.:	50 x 2.1 mm 00B-4752-AN	
1	Take 1 g homogenia	zed sample and add into 50 mL centrifuge tube.	Mobile Phase:	A: 5 mM Amm B: Acetonitrile	
2	Add 10 mL acetonit	rile and 10 mL Water.	Gradient:	Time (min)	
3	Add salts in salt pac <i>cAution</i> : Exothermic React <i>NOTE</i> : Salt pack will clump (v violently until homogenous an	kets (4 g Magnesium Sulfate and 1 g Sodium Chloride). on. ortoxing is not violent enough to break up clumps), specifically shake the sample d then vortex for 30 sec.		0 0.5 3	
4	Centrifuge until ther solids layers.	e is distinct separation between Acetonitrile, Water and		3.1 4	
5	Transfer 200 µL to a clean Acetonitrile ar	autosampler vial for LC-MS/MS analysis, or remove 1 mL ad proceed to dSPE Protocol.	Flow Rate:	0.55 mL/min	
roQ QuEChERS dSPE Kit (part No.: KS0-8920)			Injection:	5 µL	
7	Transfer 1 mL of Ac Sulfate and 50 mg F	fer 1 mL of Acetonitrile and add to dSPE kit tube (150 mg Magnesium te and 50 mg Primary/Secondary Amine).			
8	Vortex for 30 secon	ds and centrifuge.	Detector:	Agilent 6460 (
9	Remove 1 mL clean Acetonitrile and analyze or proceed to SPE Protocol.		Analytes.	2. PFPeA	
SPE usi	ng Strata™-X-AW 200		3. PFBA 4. PFHxA		
10	Condition:	3 washes of 2 mL 0.3 % Ammonium Hydroxide / Acetonitrile.		5. PFHpA 6. PFHxS	
11	Equilibrate:	3 mL Water.		7. 6:2 FTS	
12	Load:	About 15 mL of diluted QuEChERS extract.		9. PFHpS	
13	Wash:	5 mL Water.		10. PFOS 11. PFNA	
14	Elute:	4 mL 0.3 % Ammonium Hydroxide / Acetonitrile.		12. FOSA	
15	Dry:	Evaporate to near dryness and reconstitute to 500 $\mu\text{L}.$			
16	Transfer to autosam	pler vial and LC-MS/MS analysis.			

Column: Luna Omega 1.6 µm PS C18 ensions: 50 x 2.1 mm Part No.: 00B-4752-AN bile Phase: A: 5 mM Ammonium Acetate in Water B: Acetonitrile Gradient: Time (min) %**B** 0.5 3.1 100 100 Flow Rate: 0.55 mL/min Injection: 5 µL mperature: 40 °C System: Agilent 1290 Detector: Agilent 6460 QQQ Analytes: 1. PFBA 13. MeFOSE 14. 8:2 FTS 2. PFPeA 15. MeFOSA 16. PFDA 3. PFBA 4. PFHxA 5. PFHpA 17. EtFOSE 6. PFHxS 18. EtFOSA 7. 6:2 FTS 19. PFDS 8. PFOA 9. PFHpS 20. PFUdA 21. PFDoA 10. PFOS 11. PFNA 22. PFTrDA 23. PFTeDA 12 FOSA



Figure 3. Representative Chromatogram of a 0.05 ppb LLOQ.



The demand for PFAS analysis is not limited only to environmental samples, but also at low levels in food matrices for human consumption. Developed here is a fast and sensitive LC-MS/MS method to meet PFAS analysis needs in diverse food samples, including dairy and fish, down to low ppb levels. The use of the traditional QuEChERS technique enabled 1 ng/g sensitivities, and an additional (optional) solid phase extraction can help bring this down to 0.1 ng/g and ensure the optimum effectiveness of LC-MS/MS analysis.

Conclusion

Even with decreased use of PFAS on commercial products, PFAS contamination will persist throughout the environment doe to their bio-accumulative properties. This, demand for PFAS analysis is increasing and is not limited only to environmental samples such as water and soil. They are also in demand for low level testing in food matrices for human consumption and supply chain characterization. Presented here are three fast and sensitive LC-MS/MS methods developed to meet PFAS analysis needs in diverse sample matrices including water, sediments, and dairy and fish down to low ppb levels. The use of the QuEChERS technique along with further clean-up using SPE is an effective extraction and clean-up procedure to ensure the optimum effectiveness of LC-MS/MS analysis that may be adapted for most laboratory and workflow needs.