

Column Bed Stability During Routine Size Exclusion Chromatography

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Overview

Size Exclusion Chromatography (SEC) is a technique to separate large molecules such as proteins and polymers based on their size in solution. SEC methods are commonly used for routine testing application, most notably for the quantitation of aggregates in a biotherapeutic sample.

The primary failure mode for size exclusion columns is column voiding, wherein the column packed bed compresses and collapses. Column voids can occur because of spikes in backpressure, switching mobile phase (resulting in changes in solvent viscosity), or suboptimal method/sequence start up and shut down.

To demonstrate column method robustness against column voiding during routine testing, a 4.6 mm internal diameter (ID) by 150 mm length column was packed with sub-2 μm , 200 \AA media and was run with a viscous mobile phase containing 10 % isopropanol. To further stress the column packed bed, flow rate was modulated between 0.35 mL/min and 0.45 mL/min, the latter resulting in a backpressure of roughly 320 bar. Finally, flow was intermittently stopped for 8 hours to replicate the system start up and shut down which can also result in chromatographic bed collapse. Overall, there was > 100 hours of run time, and 226 injections were performed. A summary of the sequence is indicated below. The robustness sequence was to be

LC Conditions

Column: Biozen™ 1.8 μm dSEC-2, 200 \AA

Part No.: [00F-4787-E0](#)

Dimensions: 150 x 4.6 mm

Mobile Phase: 0.1 M Sodium Phosphate, pH 6.8 + 10 % Isopropanol

Flow Rate: As indicated in Table 1

Injection Volume: 10 μL

Temperature: 25 °C

Detection: UV @ 280 nm

Sample: Uridine

run until column failure, which was defined as a drop of 25 % efficiency for uridine, a small molecule which elutes in the total permeation volume.

Upon completion of the sequence, column efficiency showed a nominal drop in uridine efficiency (37231 N to 36257 N, or a 2.6 % drop). It is important to note that no protein samples were injected during the sequence, as the intent of the experiment was to confirm column bed stability. Additionally, peak shape and efficiency of uridine may not be representative of the performance of protein samples. Finally, because column failure was not reached, it is unclear as to when the 25% efficiency benchmark would have been reached.

Table 1. Experimental Conditions and Sequence for SEC Column Robustness

Sequence	Mobile Phase	Flow Rate (mL/min)	Method Run Time (min)	Number of Injections
1	0.1 M Phosphate Buffer, pH 6.8 + 10 % IPA	0.35	6.5	4
2	0.1 M Phosphate Buffer, pH 6.8 + 10 % IPA	0.45	6.5	4
3	0.1 M Phosphate Buffer, pH 6.8 + 10 % IPA	0.35	6.5	3
4	0.1 M Phosphate Buffer, pH 6.8 + 10 % IPA	0.35	90	1
5	N/A (no system flow)	0	480	0
Sequence repeated until 100 hours of run time was exceeded				



Figure 1. Column Efficiency (N) For Uridine (0.35 mL/min Method)

Column efficiency for uridine over hours of run time. Sequence as shown in Table 1. Only a 2.6 % drop in efficiency is observed over the course of the sequence.

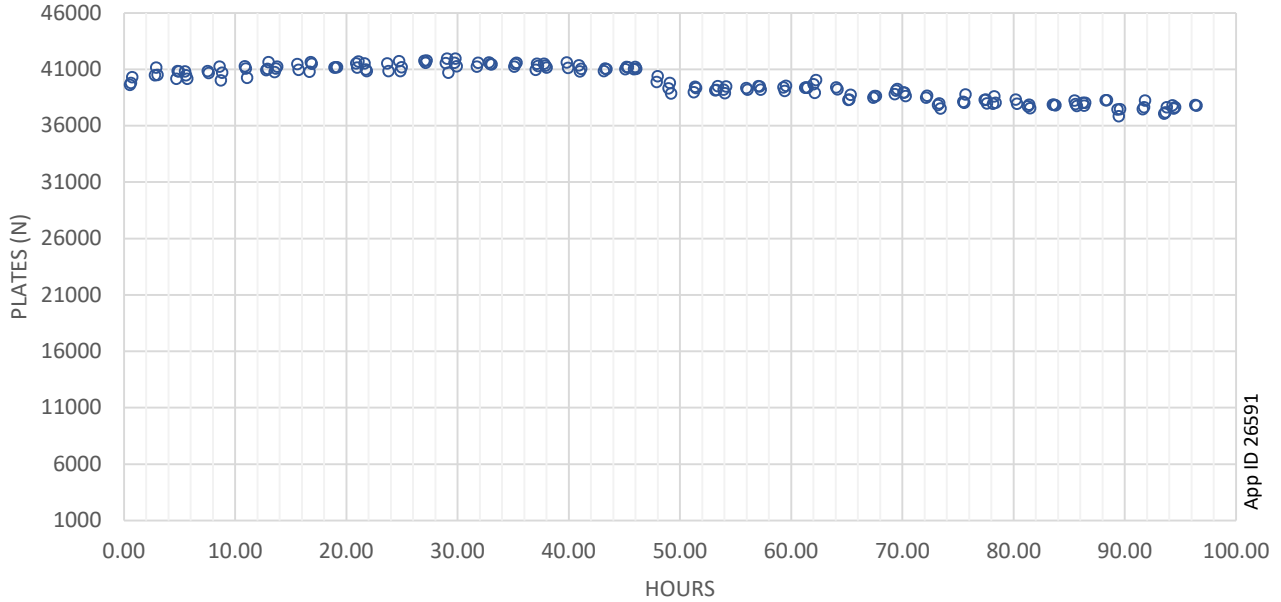
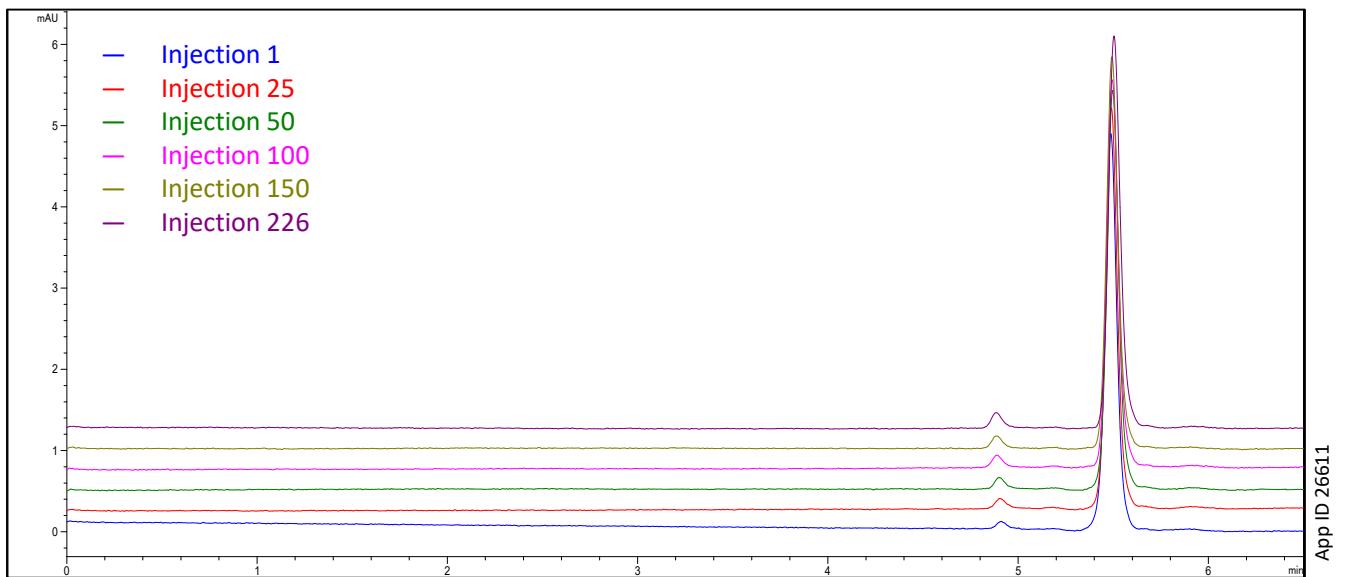


Figure 2. Chromatographic Overlay of Uridine Peak

Overlay of uridine chromatograms. Nominal changes in plates (37 k to 36 k) and retention time (5.4 min to 5.5 min) were observed through the assessment of bed stability during the sequence.



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