

## The Effect of Column Internal Diameter for Robust Size Exclusion Chromatography Methods

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### Overview

Size exclusion chromatography (SEC) is one of the primary methods for the quantitation of aggregates in a protein therapeutic. Because of its ease of use and method transferability, analytical SEC is used throughout the biotherapeutic development process.

Recently, the use of sub-2  $\mu\text{m}$  particles for SEC have expanded analytical method for the characterization of biologics. This is especially noteworthy with monoclonal antibodies (mAbs), wherein methods developed on ultra high-performance SEC (UHP-SEC) columns can separate not only aggregates, but also fragments. However, system limitations in downstream labs may not be amenable to the sub-2  $\mu\text{m}$  SEC columns which demand HPLC systems with minimal dwell volumes. Further, because they might be more prone to clogging, the UHP-SEC column format may not be the most practical choice.

A compromise to the sub-2  $\mu\text{m}$  particle would be to utilize a 3 or 5  $\mu\text{m}$  particle. Larger particles typically employ the traditional 7.8 mm internal diameter (ID), as larger IDs are more resistant to the diffusion or peak broadening associated with system dwell volume. SEC, being both isocratic and non-adsorptive, will undergo significant peak broadening with any extra column volume.

Some labs consider using the larger particle size with a smaller ID. This may be to save on mobile phase, as the 7.8 mm ID will require 3x the mobile phase as a 4.6 mm ID would. However, because the 4.6 mm ID format will be more prone to extra column volume, a performance decrease may be observed. Here we compare aggregate analysis results for a monoclonal antibody.

Figures 1 and 2 show the separation of Trastuzumab using standard mobile phase conditions. Although percent purity by peak areas and resolution of monomer/aggregate are relatively similar, the

separation of fragment, especially the post peak, is only observed with the 7.8 mm ID. Chromatographic performance is summarized in **Table 1**.

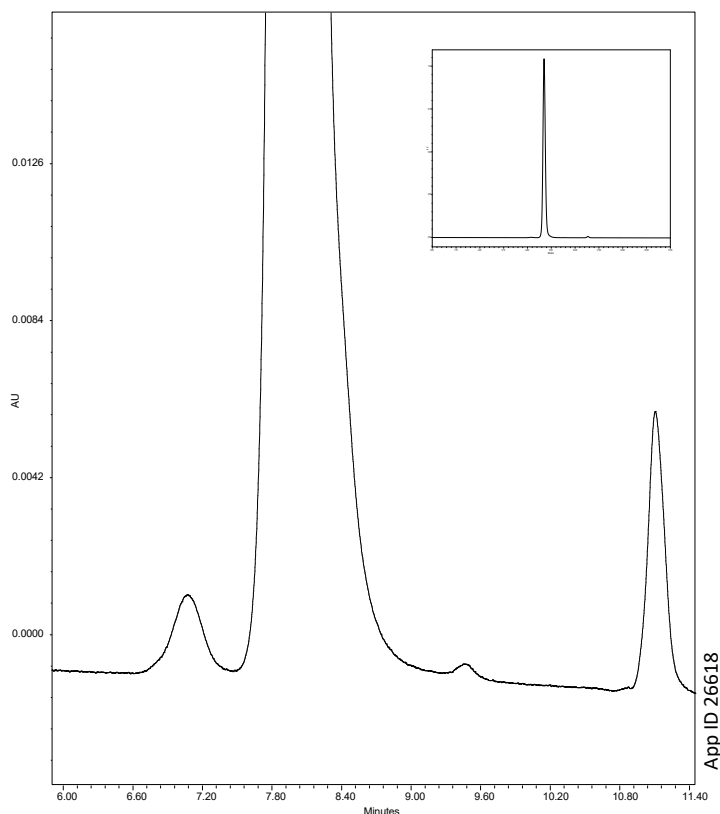
It is important to note that the concentration and injection volume are identical with both columns. Based on similar peak widths of the 4.6 mm ID, the effect here is likely not due to overloading. Rather, because the 7.8 mm ID is more resistant to extra column volume, chromatographic performance will be superior. Even though the system used for this comparison was a relatively low dwell volume system (< 400  $\mu\text{L}$ ), this is still enough to impact the SEC separation.

The downside to the 7.8 mm ID is the need for more sample and mobile phase. However, if robustness and method transferability is prioritized, the 7.8 mm ID should be considered.

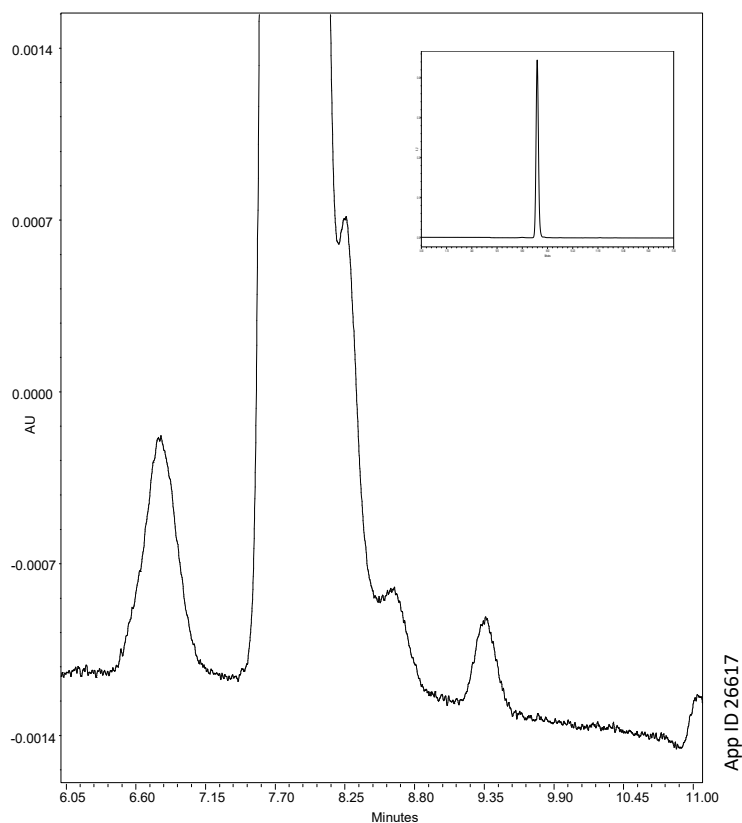
### LC Conditions

<b>Column:</b>	Biozen™ 3 $\mu\text{m}$ dSEC-2, 200 Å
<b>Dimension:</b>	300 x 7.8 mm 300 x 4.6 mm
<b>Part No.:</b>	<a href="#">00H-4788-K0</a> <a href="#">00H-4788-E0</a>
<b>Mobile Phase:</b>	50 mM Sodium Phosphate + 300 mM NaCl, pH 6.8
<b>Flow Rate:</b>	1.0 mL/min (7.8 mm ID) 0.35 mL/min (4.6 mm ID)
<b>Injection Volume:</b>	10 $\mu\text{L}$
<b>Temperature:</b>	30 °C
<b>Detection:</b>	UV @ 280 nm
<b>System:</b>	Waters® UPLC®
<b>Sample:</b>	Trastuzumab, 10 mg/mL

**Figure 1. SEC Profile for Trastuzumab (4.6 mm ID)**



**Figure 2. SEC Profile for Trastuzumab (7.8 mm ID)**



**Table 1. Chromatographic Performance Comparison, 4.6 vs 7.8 mm ID**

4.6 mm ID (Figure 1)					
Name	Retention Time	% Area	USP Resolution	USP Tailing	USP Plate Count
HMWS	7.07	0.27	-	1.05	3623
Monomer	7.984	99.3	2.36	1.1	10651
Fragment	11.107	0.43	10.56	1.15	26362
7.8 mm ID (Figure 2)					
Name	Retention Time	% Area	USP Resolution	USP Tailing	USP Plate Count
HMWS	6.799	0.18	-	1.29	6325
Monomer	7.804	99.73	2.86	1.09	11052
Fragment	12.049	0.09	15.22	1	36342



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