

New Analytical Frontiers

Perfluoroalkyl Substances (PFAS) Testing Guide



New Analytical Frontiers

The number of PFAS compounds found on current analyte lists represents less than 1 % of the potential environmental contaminants that could be contributed by this large class of compounds. However, researchers continue to identify additional PFAS compounds with potential human health and environmental impacts, thereby increasing the scope of the problem. Consequently, it is inevitable that PFAS analyte lists will continue to grow, and, future analytical challenges - sample preparation, chromatography and mass spectrometry - will become more complex and difficult to overcome. Therefore, we close this PFAS Guide with two visionary technical notes which propose new analytical approaches that will help meet the evolving PFAS challenge.



1. pH-Variable LC Mobile Phase Gradient

PFAS Analysis Based Upon a pH-Variable LC Mobile Phase Gradient

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Introduction

Polyfluoroalkyl substances (PFAS) have been an environmental concern ever since the 1970s when initial reports of potential adverse health effects first came to light. While the analysis of PFAS compounds has been ongoing for some time in academia, they are a fairly recent addition to the suite of analyses commonly performed by commercial environmental laboratories. The only official methods for the analysis of PFAS in drinking water are EPA 537/537.1 and EPA 533 and there are currently no official methods for the analysis of PFAS in complex environmental matrices such as Wastewater, Sediment, and Soil. Although ASTM has released methods for the analysis of PFAS in complex matrices (ASTM D7979 and D7968), they have not gained widespread use within the environmental testing community. As PFAS analyte lists continue to expand and matrices become ever more complex, we anticipate the need for a scalable analytical framework that will enable the development of analytical methods for a wider range of PFAS compounds and matrices. In this Technical Note we present such a framework, based upon the use of a variable pH mobile phase gradient, which could facilitate the expansion of PFAS analyte lists beyond those in common use today.



Method Limitations

Most PFAS methods in use today employ an ammonium acetate (NH_4OAc) mobile phase at a pH of 7 and with a concentration between 2 and 20mM. Although EPA methods 537.1 and 533 both specify 20mM NH_4OAc , EPA's method flexibility criteria allow for the use of alternative mobile phases (1, 2). This allowance is useful in pursuing potentially better eluent systems and allowing the analyst to run various PFAS methods on the same instrument using the same column and similar mobile phase. The benefit of changing the eluent system is the ability to change analyte selectivity and potentially analyte resolution. Selectivity differences can also be useful when trying to discriminate analytes from matrix interferences. However, the drawback to changing eluent systems is that it takes time and can create other issues associated with differing mobile phase composition.

Recently introduced regulations in California (3) have significantly expanded the PFAS target analyte list to include compounds such as PFBA, PFMBA, PFHxDA and PFOcDA, which have very large differences in hydrophobicity. This presents a significant analytical challenge because PFHxDA (C16) and PFOcDA (C18) are very hydrophobic with limited solubility in water. The predicted solubility of PFOA (C8) and PFOcDA (C18) are 480,000 and 0.00047ng/L respectively, using the WS-KOWIN from the USEPA EPISuite Software (4).

In addition, chromatographic analysis of PFBA in an extract that is > 90 % organic results in poor peak shape for this early eluting compound. Most methods that can successfully analyze for PFBA are either direct injection (100 % water), a 1:1 water-methanol dilution or have at least 20 % water in the extract (EPA 533). Some methods (ASTM D7979, D7968 and EPA 8327) add acetic acid to the extract to help improve the peak shape of PFBA. However, this results in poorer chromatographic performance for the longer chain PFHxDA (C16) and PFOcDA (C18).

A New Strategy

In recognition of these limitations, we have pursued a new chromatographic strategy using a 100 % organic system (for long chain PFAS solubility) and variable mobile phase pH to provide good chromatography for PFBA and other early eluting PFAS compounds. By staying within the confines of the NH_4OAc mobile phase composition but employing pH as a variable, one can realize the potential advantages mobile phase variation allowed by EPA while avoiding the primary disadvantages. This approach could be useful in overcoming the difficulty of expanding the analyte lists of the existing PFAS methods to incorporate both the hydrophilic shorter chain compounds and the extremely hydrophobic longer chain compounds.

Technical Approach

This work specifically focused on a secondary chemical characteristic of most PFAS compounds: the hydrophilic or polar functional head of the molecule which are either carboxylic or sulfonic acids which can be charged or neutral, depending on the pH of the eluent. Chromatographers can take advantage of secondary interactions by employing a mobile phase in which a pH gradient is performed, i.e. changing the pH of the mobile phase over time. Mobile phase pH becomes important when analytes contain acidic, basic or both functional groups. The mobile phase pH determines the charge state (protonation state) of the analyte and thereby influences its interactions with the mobile and stationary phase. This technique allows for more control of the ionic interactions between the PFAS analytes within a column's stationary phase and the mobile phase. This is analogous to the WAX SPE technique used in EPA method 533, wherein the ion exchange mechanism allows for stronger interaction with the shorter-chain PFAS compounds than does the styrenedivinylbenzene (SVDB) SPE sorbent used in method 537.1 which operates primarily in a reversed phase mode. Shorter chain PFAS compounds have a lower degree of binding ability due to their shorter chain length and thus often pass through, owing to binding mechanisms that rely exclusively or primarily on a reversed phase interaction.

1. pH-Variable LC Mobile Phase Gradient (continued)

In this new technique, the mobile phase at the beginning of the run has a low pH (~ pH 3.9) and changes over time to a higher pH (~ pH 9.3). This protonates or deprotonates the functional heads of the various PFAS compounds over time, depending upon the pKa of the functional group. This correspondingly changes the elution profile for the separation, in terms of both relative and absolute retention times. In principle, the protonation of short-chain, anionic PFAS will lead to greater retention, while the deprotonation of the later-eluting, long-chain PFAS may lead to less retention, thereby compressing the chromatogram. This will lead to less suppression from non-retained interferences, and shorter run times, allowing greater sample throughput. Separating interferences from early eluting analytes is particularly important when there is only one sensitive MRM transition available, as in the case of PFBA and PFPeA. It is reasonable to think that these orthogonal retention mechanisms (hydrophobicity vs. ionizability or pKa) could offer greater opportunity to resolve complex PFAS mixtures. This Technical Note provides an illustration of the potential power of this approach.

Experimental Conditions

Instrumentation and Consumables. All PFAS analyses were performed on an Agilent® 1100 HPLC with a Thermo Scientific® TSQ Vantage triple quadrupole mass spectrometer. All samples were prepared using a Phenomenex Strata®-X-AW 200mg 33µm in a 6cc format (pn: [8B-S038-FCH](#)). The LC column employed was a Phenomenex Kinetex™ C18 EVO 5µm 100 x 2.1 mm (pn: [00B-4633-AN](#)).

Reagent Preparation. Eluents: (1A) Ammonium Acetate (NH₄OAc) was prepared at 20mM by dissolving 1.54g NH₄OAc into 1.0 L of water. LC-MS methanol (MeOH) was used for (1B). Acetic acid (HOAc) was prepared at 20mM by diluting 1.22 mL of glacial acetic acid into 1.0 L of water (2A). Basic methanol was prepared by diluting 1.46 mL of concentrated Ammonium Hydroxide (NH₄OH) into 1.0 L of LC-MS methanol. Reference materials were purchased from Wellington Labs (Guelph, Canada) and diluted into LC-MS methanol for analysis.

Mass Spectrometer Operating Conditions: The capillary and vaporizer temperature were 250 °C and 300 °C respectively. The sheath and aux gas were held at 40 arb and 50 arb respectively. The ESI voltages for positive and negative mode were +3.0/-2.5add spacekV. See Appendix 1 for MS/MS Parameters.

LC Operating Conditions: A moderate organic gradient profile was used in both analyses being compared. The only difference between the two LC systems was the pH modifiers that were used in the aqueous and organic eluents. To illustrate the effect of improved peak shape and selectivity differences solely due to the pH modifiers, the times used to change from aqueous to high organic were identical.

Results and Discussion

Although it is difficult to determine the actual pH in any eluent system especially in the presence of methanol and a particular stationary phase, this was estimated in an offline experiment. In order to ascertain the pH change as 20mM HOAc mixes with the 25mM NH₄OH, the pH was measured offline for different mixture ratios of this binary system. The measured pH values are shown in **Table 3**. Based on this data, it is estimated that the gradient pH elution profile has a pH no wider than 3.9 and 9.3 from start to finish respectively.

Table 1.
LC Conditions (neutral, pH=7)

	20 mM NH ₄ OAc	MeOH
Time	% A	% B
0.00	95	5
1.20	55	45
3.60	35	65
11.00	10	90
13.00	10	90
13.01	95	5
17.00	95	5

Table 2.
LC Conditions (gradient pH)

	20 mM HOAc	25 mM NH ₄ OH in MeOH
Time	% A	% B
0.00	95	5
1.20	55	45
3.60	35	65
11.00	10	90
13.00	10	90
13.01	95	5
17.00	95	5

Table 3.
Measured pH of a Binary Mixture of Eluents

20 mM HOAc	25 mM NH ₄ OH in MeOH	
% A	% B	Actual pH
100	0	3.62
95	5	3.86
90	10	4.17
80	20	4.55
70	30	5.14
60	40	5.77
50	50	6.45
40	60	7.13
35	65	8.15
30	70	8.52
20	80	8.98
10	90	9.33
0	99.5	10.25

1. pH-Variable LC Mobile Phase Gradient (continued)

One of the first notable improvements using the new gradient pH upon injecting an extract containing PFAS in 100% methanol is that the peak shape for PFBA is drastically improved due to shifting the equilibrium of unprotonated PFBA to a protonated form. Protonated PFBA will interact with the nonpolar stationary phase much more than the mobile phase causing increased retention and a better focused peak. This is illustrated in **Figures 1 and 2**; PFBA (light blue). Under the commonly used eluent system of 20mM NH₄OAc, PFBA and PFMPA exhibit severe fronting in 100% methanol (required for PFODA solubility). However, using the gradient pH profile, these peaks are focused much better on the column.

Additionally, the latest eluters (PFTrA, PFTeDA, PFHxDA and PFODA) not only elute early, but the peak height is noticeably higher. The increase in height would improve detection limit with a greater s/n. This indicates that NH₄OH, which increases in concentration as the organic (methanol) gradient progresses, is affecting analyte retention by shifting their equilibrium to a deprotonated anion since the anions favor interactions with the mobile phase and the neutral analyte favors interaction with the stationary phase. In fact, the NH₄OH must be present in slightly higher molar concentration than the HOAc in order to move the pH into the slightly basic range.

Figure 1.
Chromatogram of 48 PFAS using 20mM NH₄OAc (pH=7)

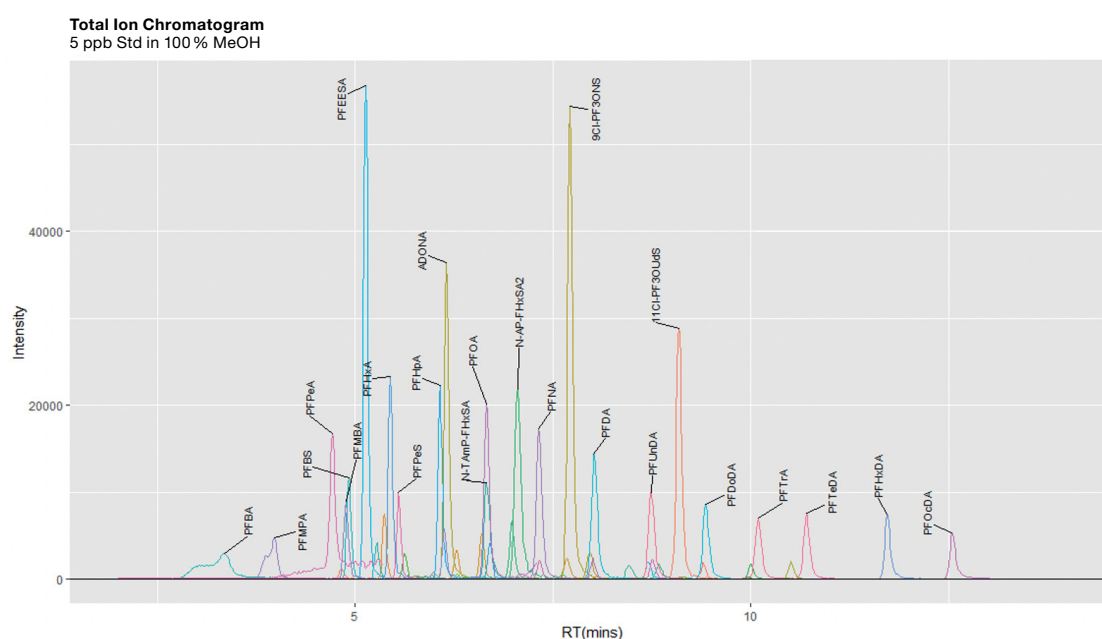
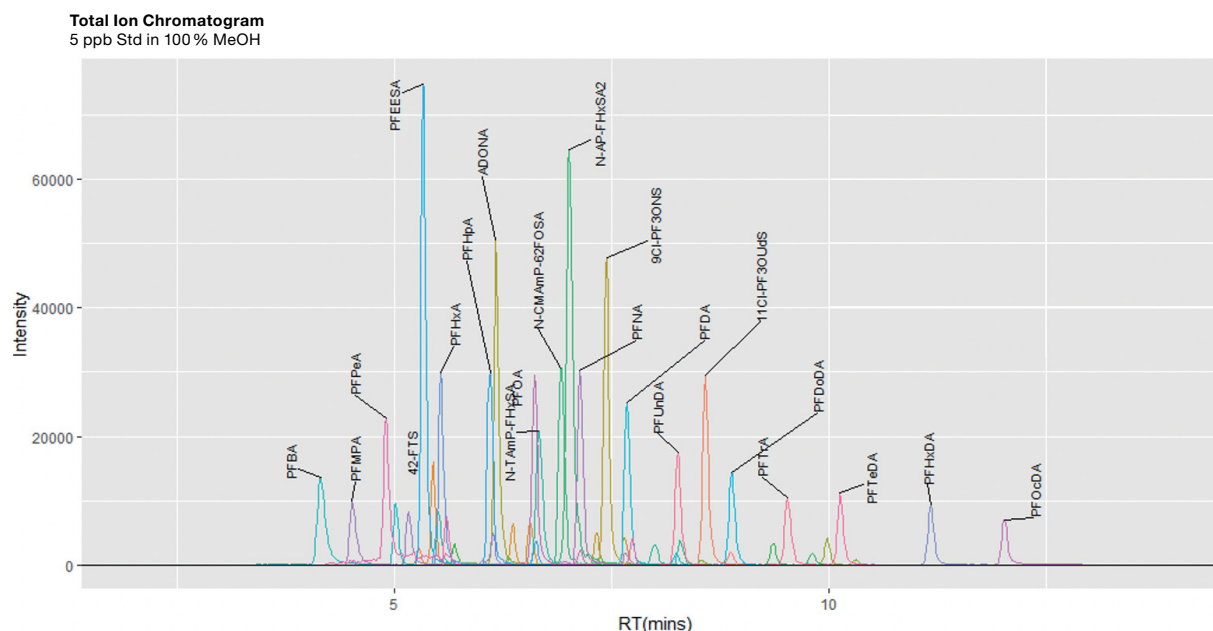


Figure 2.
Chromatogram of 48 PFAS using 20mM HOAc and 25mM NH₄OH (varied pH from 3.9 to 9.3)



1. pH-Variable LC Mobile Phase Gradient (continued)

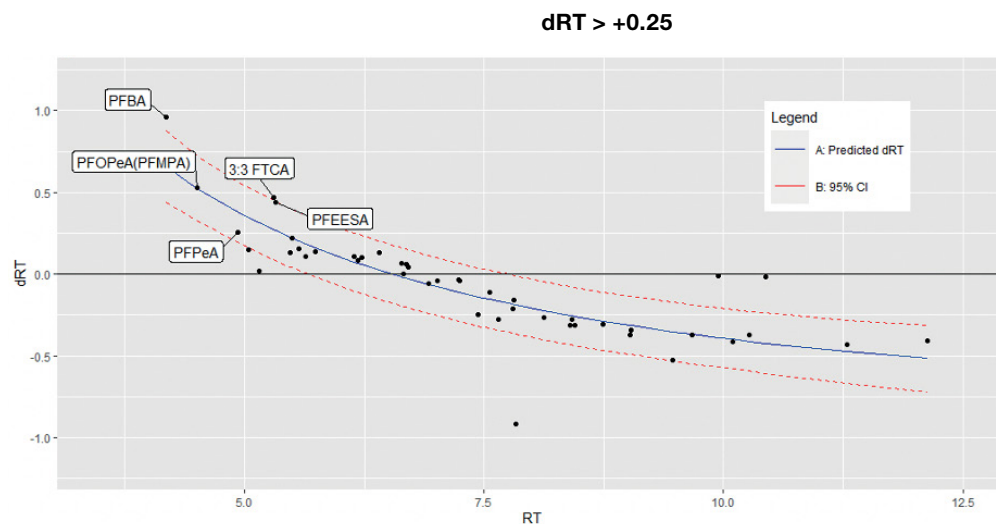
The selectivity of these two mobile phase systems was further investigated to see how they affect different PFAS compounds varying in chain length.

Upon close examination of the ΔRT data there were certain analytes (e.g PFOSA) that indicated possible differences in selectivity. In order to evaluate significant selectivity differences between the two eluent systems that were not obvious, a statistical approach was used. This is necessary because not every slight change in RT or resolution may be significant. First, a least squares regression was performed on the ΔRT as a function of RT of the new method. The equation that was used to model the change in the two systems is listed in equation (1) where a, b, c are the coefficients for the intercept, linear term, and inverse term respectively:

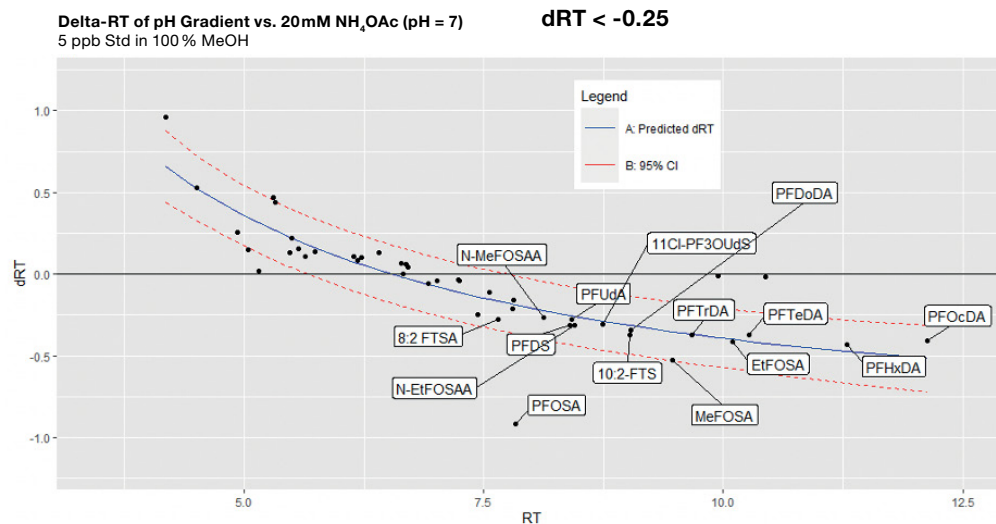
$$\Delta RT = a + b \cdot RT + c \cdot \frac{1}{RT} \quad (1)$$

To validate the regression model and the prediction interval of significance at 95 %, a Global Validation of Linear Models Assumptions (GVLMA) was used (5) The plots in **Figure 3** highlight the most important aspects of the advantages of this new system. These are increased retention for early eluters (3a), decreased elution for late eluters (3b), and significant selectivity differences (3c). To evaluate significant differences, the x-axis shows the retention time (RT) for the new mobile phase and the y-axis shows the ΔRT relative to the neutral ammonium acetate mobile phase.

Figure 3.
Notable Mobile Phase Elution Changes
a) PFAS Analytes with Increased Retention



b) PFAS Analytes with Decreased Retention



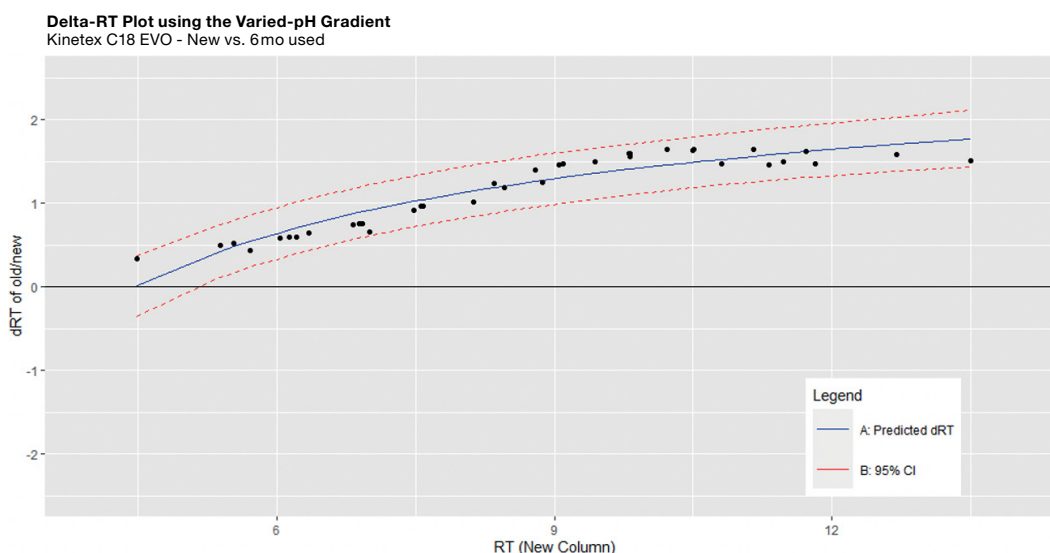
1. pH-Variable LC Mobile Phase Gradient (continued)

It is also worth noting that this new eluent system also has an effect on sensitivity for certain compounds. Specifically, N-TAmP-FHxSA, N-CMAmP-62FOSA, and N-AP-FHxSA2 (which are detected in ESI+) had an increase in response more than 2x in the new pH gradient eluent system (Figure 1-2).

Lastly, the robustness of the stationary phase was examined by evaluating a “well used” LC column versus a brand new column. The “well used” column had been used to analyze thousands of samples over approximately six months. This included drinking water extracts as well as non-potable aqueous and soil extracts. The Kinetex™ EVO C18 showed reasonable robustness and, although some retention is lost over time, there was no significant ($P < 0.05$) selectivity difference observed. Again, the GVLMA cross-validation was used (Figure 4) to detect significant elution order changes (ie: all analytes had statistically the same elution order) although “absolute” elution order was different in some cases.



Figure 4.
Retention Difference of New vs Used column Under Varied pH conditions



Conclusion

The objective of using a pH gradient mobile phase for PFAS analysis is that it allows the analyst to widen the scope of analyte chemistry to properly chromatograph short-chain and long-chain PFAS in 100% organic extracts as well as change the selectivity of the method. This holds true for any analyte panel outside the scope of method EPA 537.1 and EPA 533, in that the absolute and relative retention of some analytes are different than when using a standard organic gradient with ammonium acetate (NH_4OAc).

Additionally, this solution may provide the ability to move certain peaks away from interferences and high ion suppression zones at the beginning of the chromatographic run. It may also allow for the inclusion of other PFAS analytes with a minimal redevelopment and optimization. The pH gradient method shows excellent robustness and reproducibility, with stable PFAS analyte retention times, even when using different columns, systems, and analysts. The changes in retention times (both absolute and relative) offer another tool for more complex PFAS mixtures - either those with more PFAS analytes or from working with dirtier matrices.

Moving forward, this promising mobile phase gradient approach could be combined with work investigating alternative HPLC stationary phases to determine optimal conditions for PFAS panels that are much broader in scope and chemistry. In principle, this approach should allow the separation of an even wider class of PFAS including non-volatile short-chain PFAS. Preliminary data suggest that the use of Formic acid (ie: 25 mM HOFo) instead of 25 mM HOAc can drop the pH slightly lower; closer to pH=3. This has the benefit of increased retention for TFA, TFMS, and PFPrA in extracts that are 100% methanol.

1. pH-Variable LC Mobile Phase Gradient (continued)

Appendix 1. Instrumental Conditions for MS/MS Analysis and RT Data

Analyte	Precursor	Product	CE	S-Lens	Polarity	Retention Time Data	
						Gradient pH	Constant pH=7
PFBA	213	169	9	35	-	4.18	3.22
PFMPA	229	85	12	35	-	4.51	3.98
PFPeA	263	219	9	38	-	4.93	4.68
3:3-FTCA	241	177	8	37	-	5.30	4.83
PFEESA	315	135	23	90	-	5.32	4.88
PFBS	299	80	36	100	-	5.04	4.89
PFMBA	279	85	12	40	-	5.15	5.13
NFHDA	295	201	10	33	-	5.50	5.28
4:2-FTS	327	307	20	110	-	5.48	5.35
PFHxA	313	269	9	47	-	5.57	5.42
PFPeS	349	80	41	100	-	5.64	5.53
HFPO-DA	285	169	8	37	-	5.74	5.60
PFHpA	363	319	9	56	-	6.15	6.04
PFHxS	399	80	44	120	-	6.18	6.10
ADONA	377	251	11	60	-	6.22	6.12
5:3-FTCA	341	237	13	57	-	6.41	6.28
6:2-FTS	427	407	22	130	-	6.64	6.57
PFOA	413	369	9	62	-	6.69	6.63
N-TAmP-FHxSA	499.1	60	37	140	+	6.66	6.66
PFHpS	449	80	46	110	-	6.71	6.66
N-CMamP-6:2FOSA	571.1	440	31	140	+	6.92	6.98
N-AP-FHxSA	485.1	85	34	130	+	7.01	7.05
PFNA	463	419	10	65	-	7.24	7.27
PFOS	499	80	46	105	-	7.25	7.29
9Cl-PF3ONS	530.9	351	28	120	-	7.56	7.67
7:3FTCA	441	337	11	70	-	7.44	7.68
8:2-FTS	527	507	27	130	-	7.65	7.93
PFDA	513	469	10	75	-	7.81	7.97
PFNS	549	80	48	130	-	7.80	8.01
N-MeFOSAA	570	419	20	120	-	8.13	8.40
PFUnDA	563	519	10	85	-	8.42	8.69
PFDS	599	80	49	110	-	8.40	8.71
PFOSA	498	78	34	110	-	7.84	8.75
N-EtFOSAA	584	419	20	120	-	8.45	8.76
11Cl-PF3OUdS	630.9	451	30	120	-	8.74	9.05
PFDoDA	613	569	12	92	-	9.04	9.38
10:2-FTS	627	607	31	150	-	9.03	9.40
MeFOSE	616	59	15	90	-	9.95	9.96
MeFOSA	512	169	30	110	-	9.48	10.00
PFTrA	663	619	12	101	-	9.67	10.05
EtFOSE	630	59	15	91	-	10.45	10.46
EtFOSA	526	169	30	120	-	10.10	10.51
PFTeDA	713	669	12	108	-	10.27	10.64
PFHxDA	813	769	12	120	-	11.29	11.72
PF0cDA	913	869	13	140	-	12.13	12.54

References

- 1) U.S. Environmental Protection Agency, Method 537.1—Determination of Selected Per- And Polyfluorinated Alkyl Substances In Drinking Water By Solid Phase Extraction And Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS). November 2018.
- 2) U.S. Environmental Protection Agency, Method 533— Determination of Per- And Polyfluoroalkyl Substances In Drinking Water By Isotope Dilution Anion Exchange Solid Phase Extraction And Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS). November 2019.
- 3) Analytical Reporting Limits for PFAS compliance with DoD Table B-15 of QSM, Version 5.1 or later (updated 07/22/20) https://www.waterboards.ca.gov/pfas/docs/reporting_limits_dod_qsm_v5_1_or_later_july_22_2020.pdf
- 4) U.S. Environmental Protection Agency, Estimation Program Interface Suite (EPI Suite) Software, v4.11, 2000-2017. <https://www.epa.gov/tsc-screening-tools/download-epi-suite-estimation-program-interface-v411>
- 5) Peña, Edsel A., and Elizabeth H. Slate. "Global Validation of Linear Model Assumptions." *Journal of the American Statistical Association*, vol. 101, no. 473, 2006, pp. 341–54, doi:10.1198/016214505000000637.

2. Column Chemistry Considerations

Column Chemistry Considerations for Full Coverage of PFAS Analyte Ranges

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Introduction

Per- and polyfluorinated alkyl substances (PFAS) are man-made chemicals, that have been widely used since the 1940s. They have been employed in a large variety of consumer products, such as nonstick cookware, food containers, stain and water repellent fabrics, polishes, waxes, paints, and cleaning products and are now widely distributed in the global environment. A significant source of PFAS environmental contamination has been the widespread use of PFAS-containing aqueous firefighting foams (AFFF), which are known to migrate into groundwaters at airports and military bases. Further environmental exposure to PFAS comes from industrial production facilities (e.g. chrome plating, electronics, manufacturing, or oil recovery). Living organisms, including plants, animals, and humans, can accumulate PFAS compounds in their tissue, which can build up over time and impact their health.¹⁻³ A total of 9,252 PFAS are listed in EPA's most recent list of PFAS substances.⁴ However, only a handful of these, such as perfluorooctanoic acid (PFOA) and perfluorooctanesulfonic acid (PFOS), have been widely monitored in the environment or have been thoroughly studied for their toxicological effects.



Common Chromatographic Approaches

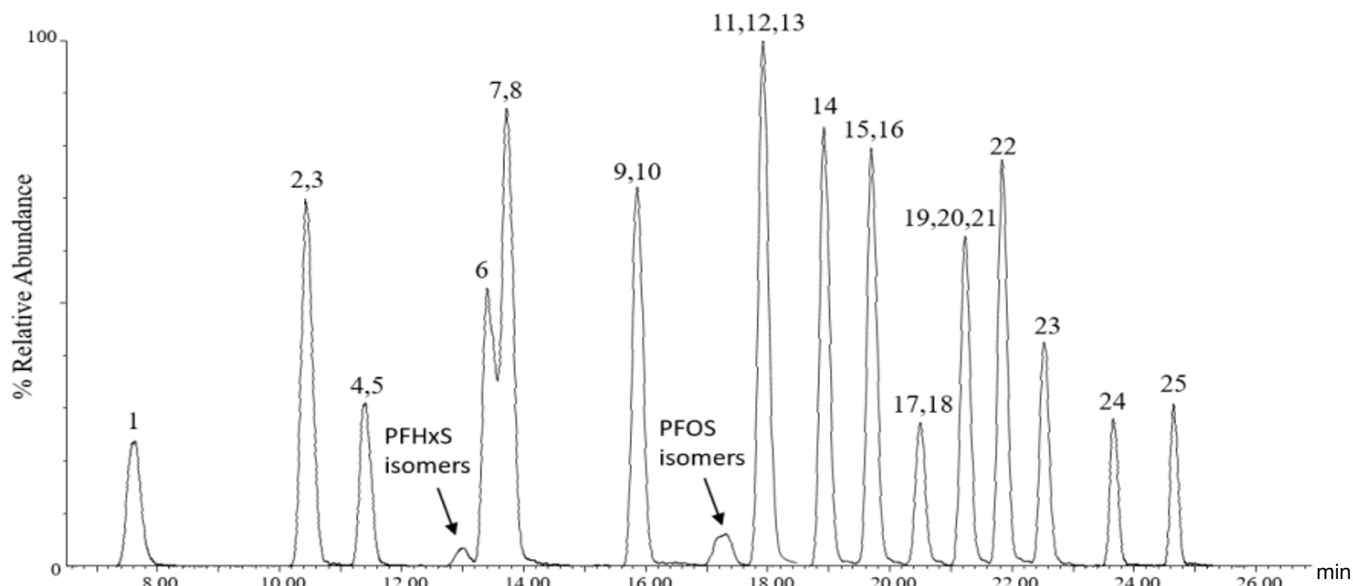
PFAS compounds are typically determined by LC-MS/MS and LC-HRMS instrumentation. The use of mass spectrometry detection has played a significant role in the quantitation of specific compounds where standards are available. Where standards are not available, the use of time of flight (TOF) and Orbitrap™ MS detectors are used to semi-quantify unknown PFAS compounds. The chromatographic separation of PFAS compounds in currently validated methods typically involves a reversed phase mechanism using a C18 or Phenyl column in an acidic-methanol eluent. For example, EPA method 537.1 uses a C18 column (5 μm, 2.1 x 150 mm C18) and EPA Method 533 was validated using a C18 Phenomenex Gemini™ column (3 μm, 2 x 50 mm). Conversely, ASTM D7979 and EPA 8327 were validated using a Phenyl-Hexyl column (1.7 μm, 2.1 x 100 mm), ISO 21675 used a C18 column (5 μm, 2 x 50 mm) and the Department of Agriculture CLG-PFAS 2.01 method used a C8 column, Phenomenex Luna™ C8(2) (3 μm, 2 x 50 mm).

PFAS Chromatographic Challenges

While these methods are generally adequate for a limited list of analytes, the large number of potential PFAS analytes that could potentially be present in a sample will inevitably challenge simple chromatographic separation approaches. This phenomenon was seen early in the development of the EPA drinking water methods. EPA 537.1 when validated, identified several overlapping peaks which can be seen in **Figure 1** as demonstrated by peaks, 2,3; 4,5; 7,8; 9,10; 11,12,13; 15,16; 17, 18; 19, 20, 21.

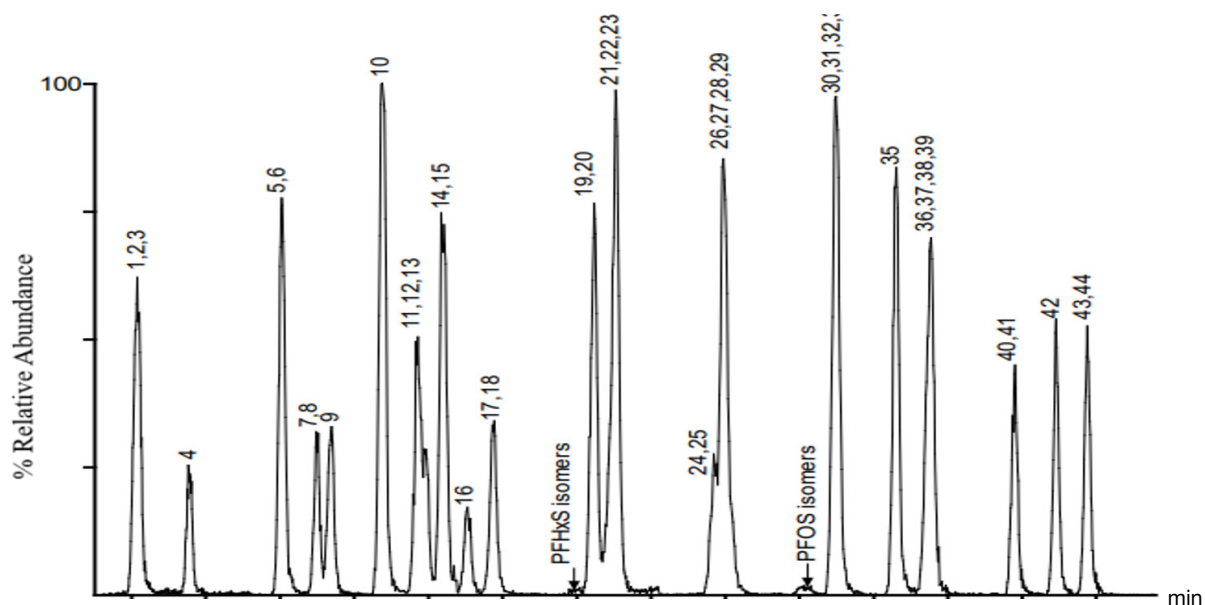
2. Column Chemistry Considerations (continued)

Figure 1.
Example chromatogram for reagent water fortified with method 537.1 analytes at 80 ng/L



Likewise, when EPA 533 was developed and validated with an expanded list of PFAS compounds, it also shows several overlapping peaks, as seen in **Figure 2**.

Figure 2.
Example chromatogram for reagent water fortified with method 533 analytes at 80 ng/L



2. Column Chemistry Considerations (continued)

Whereas many of these overlapping peaks can be successfully resolved by the mass analyzer, the potential presence of isobaric homologues and unresolved matrix interferences point to the continuing need for good chromatographic separation to assure reliable identification and quantitation. Although the problem may be manageable for today's small analyte lists, the challenge will inevitably grow as new PFAS compounds are added for investigational or regulatory purposes.

Looking to the Future

Current PFAS methods primarily rely upon C18 solid phase chemistry and simple methanol-ammonium acetate mobile phase gradients. These methods do not make full use of all the tools in the chromatographer's toolbox, nor need they, given today's limited analyte lists. However, this simple situation will inevitably change and there will be a need to develop more sophisticated chromatographic methods to tease out the subtle chemical and structural differences between closely related PFAS compounds. Chief among these will be the application of novel stationary phases and mobile phases to exploit the different interactions between closely related PFAS molecules. This Tech Note was designed to provide a vision of the potential power of such new chromatographic approaches.

Scope

In this Tech Note we will present data for a select list of PFAS compounds (Table 1) that were selected to reflect some of the chemical diversity of the PFAS universe. This color-coded grouping will be used to illustrate the differences in chromatographic retention time and elution order between various stationary phases including C8, C18, Phenyl-Hexyl, Biphenyl and F5 which can have significantly different sorptive properties. We will also examine how differing mobile phase polarity (e.g., methanol vs. acetonitrile) influences chromatographic performance for these various phases. Ideally, this information can be used to enhance chromatographic resolution as the list of PFAS compounds continues to increase. The goal is to provide insights that will allow method developers to identify useful separation strategies.

Method Variables

PFAS Chemistries

There are established, validated methods set forth by the EPA and ISO for chromatographic separation of PFAS compounds using specific types of columns and packing materials. Unfortunately, not all PFAS compounds can be separated with sufficient accuracy using these methods because of the different types of functional groups that are on different PFAS compounds. In the select list that was used, there are 5 categories of PFAS compounds as shown in Figure 3, with an example of each. Owing to the variety of functional groups that can potentially be found on PFAS compounds, there are a variety of HPLC column chemistries that could aid enhanced separation.

Figure 3.

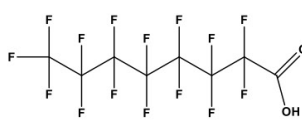
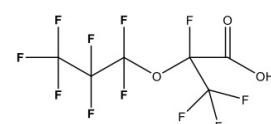
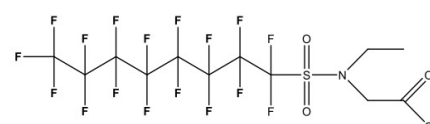
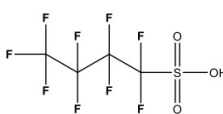
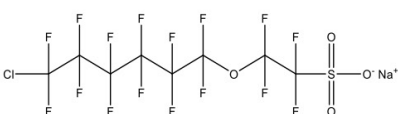
Perfluoroalkyl carboxylic acids (PFCAs)	Perfluoroalkyl ether carboxylic acids (PFECAs)	Perfluoro sulfonamide carboxylic acids (PFSACAs)
 <p>Perfluoro-n-octanoic acid</p>	 <p>Hexafluoropropylene oxide-dimer acid</p>	 <p>N-ethylperfluoro-1-octanesulfonamidoacetic acid</p>
Perfluorinated sulfonic acids	Chlorinated polyfluoroalkyl ether sulfonic acids	
 <p>Perfluorobutanesulfonic acid</p>	 <p>9-chlorohexadecafluoro-3-oxanonane-1-sulfonate</p>	

Table 1.

Chemical Name	Abbreviation
Perfluoroalkyl carboxylic acids (PFCAs)	
Perfluorohexanoic acid	PFHxA
Perfluoroheptanoic acid	PFHpA
perfluoro-n-octanoic acid	PFOA
perfluoro-n-nonanoic acid	PFNA
perfluoro-n-decanoic acid	PFDA
perfluoro-n-undecanoic acid	PFUdA
perfluoro-n-dodecanoic acid	PFDoA
perfluoro-n-tridecanoic acid	PFTrDA
perfluoro-n-tetradecanoic acid	PFTeDA
Perfluoroalkyl ether carboxylic acids (PFECAs)	
hexafluoropropylene oxide-dimer acid	HFPO-DA
dodecafluoro-3H-4,8-dioxanonanoate	NaDONA
Perfluorooctane sulfonamides and derivatives	
N-methylperfluoro-1-octanesulfonamidoacetic acid	N-MeFOSAA
N-ethylperfluoro-1-octanesulfonamidoacetic acid	N-EtFOSAA
Perfluorinated sulfonic acids (PFSAs)	
Perfluorobutanesulfonic acid	L-PFBS
perfluoro-1-hexanesulfonate	L-PFHxS
perfluoro-1-octanesulfonate	L-PFOS
Chlorinated polyfluoroalkyl ether sulfonic acids (Cl-PFESAs)	
9-chlorohexadecafluoro-3-oxanonane-1-sulfonate	9Cl-PF30NS
11-chloroicosalfluoro-3-oxaundecane-1-sulfonate	11Cl-PF30uS

2. Column Chemistry Considerations (continued)

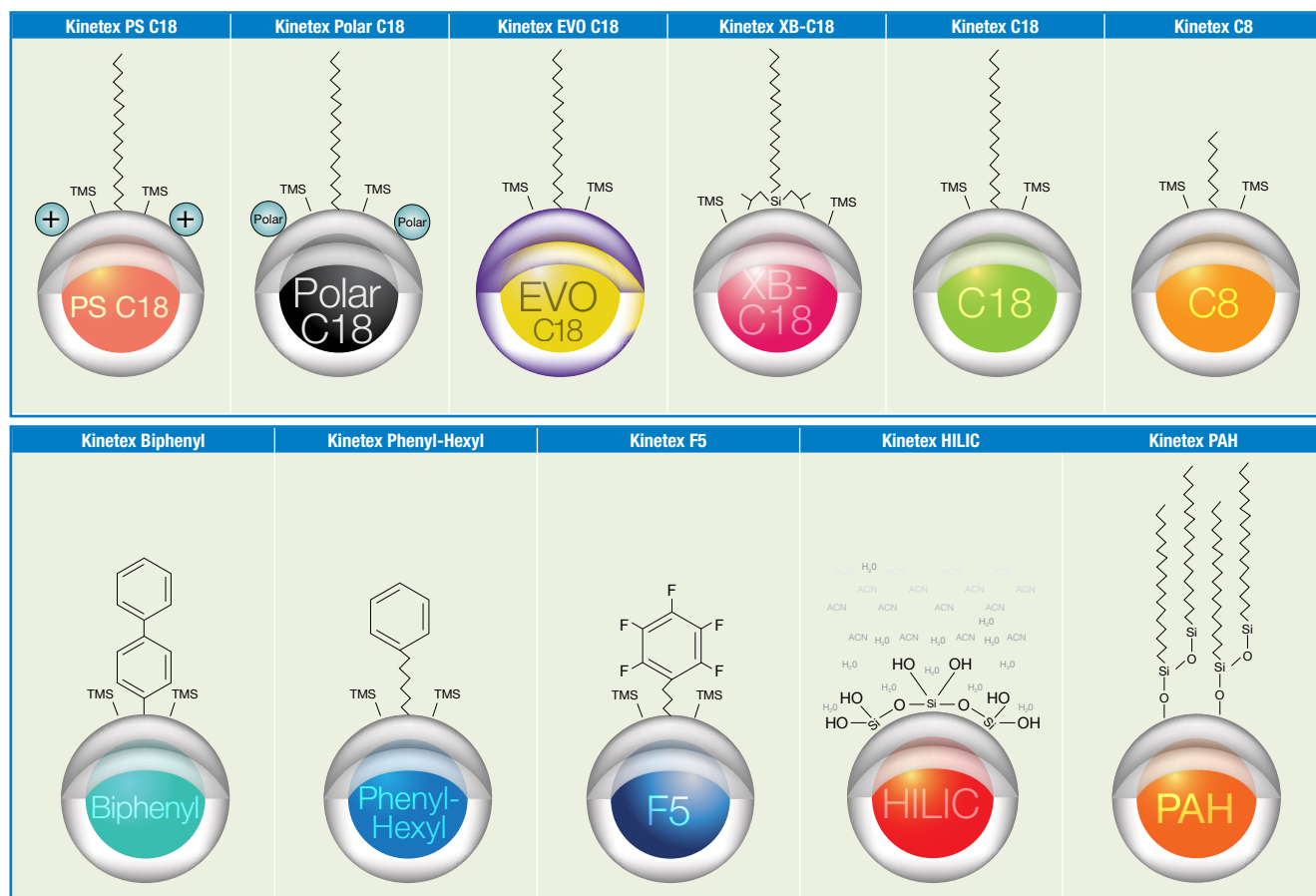
Solid Phase Chemistries

A representation of the different solid phase chemistries that are available in Phenomenex HPLC columns that could be used in PFAS separations is presented in **Figure 4**. This wide variety of ligand chemistries – combined with differences in porosity and other morphological variations – was developed to offer a wide range of variables for method development.

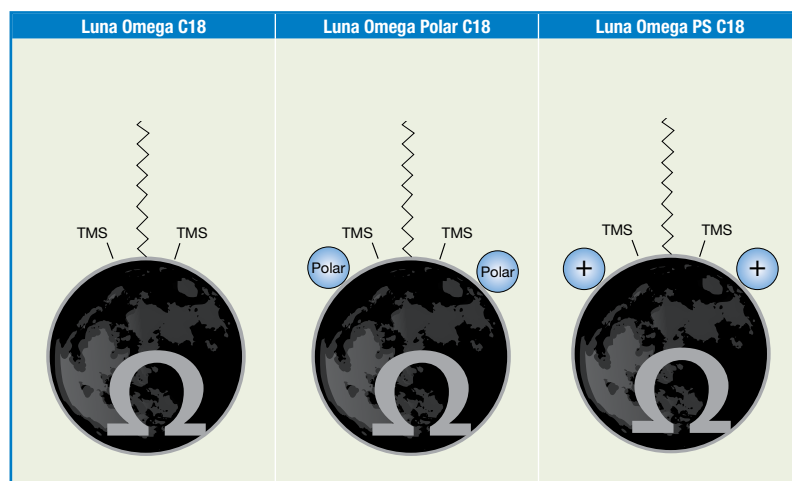
Different combinations of these variables serve to enhance the separation of polar compounds, increase surface areas, add pH stability, decrease system backpressures, etc. These, and additional column properties, provide chromatographers with a high degree of flexibility with which to tackle challenging separations.

Figure 4.
Available column chemistries appropriate for PFAS compound separation

Kinetex™ Core-Shell 1.3, 1.7, 2.6, and 5 μm



Luna™ Omega Fully Porous 1.6, 3, and 5 μm



2. Column Chemistry Considerations (continued)

Mobile Phase Chemistries

However, in addition to column selection, chromatographers can also make changes in mobile phase polarity to further enhance selectivity. For example, EPA method 533 was altered in several ways to enhance separation of the selected PFAS compounds used in the present study. In the first elution regime, the percentage of methanol was increased at run initiation and then further increased to a higher percentage than had been previously used in the published method. This decreased the overall run time but kept the percentage increase of methanol roughly the same. This elution regime will be referred to later in this Tech Note as “533 Similar” (Table 2).

In the second elution regime, acetonitrile was added to the mobile phase at a ratio of 80:20 methanol:acetonitrile to increase mobile phase polarity (but with all other factors remaining the same as in the “533 Similar” elution regime). This second elution regime will be referred to as “533 Acetonitrile Altered” (Table 3). The results from these two elution regimes will be addressed separately. Clearly, there are many other potential mobile phase variations that could be investigated. However, the two variations presented here will suffice to demonstrate the power of mobile phase polarity combined with solid phase chemistry variation to effect PFAS chromatographic behavior.

Table 2.

EPA 533 - As Published		
Time (min)	% 20 mM Ammonium Acetate	% Methanol
Initial	95	5
0.5	95	5
3	60	40
16	20	80
18	20	80
20	5	95
22	5	95
25	95	5
35	95	5

40 → 80 in 13 min 3.08 % per min

533 Similar		
Time (min)	% 20 mM Ammonium Acetate	% Methanol
Initial	55	45
15	10	90
21	10	90
21.5	55	45

40 → 90 in 15 min 3.0% per min

Table 3.

533 Similar		
Time (min)	% 20 mM Ammonium Acetate	% Methanol
Initial	55	45
15	10	90
21	10	90
21.5	55	45

533 Acetonitrile Altered		
Time (min)	% 20 mM Ammonium Acetate	% 80-20 Methanol Acetonitrile
Initial	55	45
15	10	90
21	10	90
21.5	55	45

Results and Discussion

For ease of comparison, all chromatographic data will be presented in tabular format with the chromatography columns on the left, the PFAS compounds across the top, and the specific analyte retention times under the PFAS compounds. The highlighted boxes identify two compounds that have overlapping retention times ($\Delta RT \leq 5$ seconds) and the arrows at the bottom indicate when two compounds have changed elution order. The different PFAS compound classes are represented by the colors referenced in Table 1. This representation is a more insightful way to present the data because overlaying or stacking individual chromatograms makes it very difficult to compare results across columns. The two mobile phase chemistry regimes identified above will now be discussed separately.

2. Column Chemistry Considerations (continued)

1. EPA 533 Similar

In order to determine how the selected PFAS compounds would elute and separate, seven different chromatography columns with different solid phase chemistries were examined. **Figure 5** displays columns that have C18-functionality or PAH-functionality. The PFAS elution order was generally consistent for most of the C18 phases, although specific elution times varied. The Kinetex™ PAH column demonstrated two compound functional pairs with a reverse elution order: NaDONA (a perfluoroether carboxylic acid) vis-à-vis L-PFHxS (a perfluorinated sulfonic acid) and PFUdA (a perfluoroalkyl carboxylic acid) vis-à-vis N-EtFOSSA (a perfluorooctane sulfonamide). In addition, there were slight differences in overlapping peaks amongst the various C18 phases, whereas the Kinetex PAH phase had only one overlapping pair. When compared to two C8 phases (**Figure 6**), the elution order was similar to the C18 phases, and the retention times were similar, but there were fewer overlapping peak pairs (one pair vs. 3 pairs).

However, the C8 phases also demonstrated two compound functionality pairs with a reverse order elution from the C18 phases: L-PFOS (a perfluorinated sulfonic acid) vis-à-vis PFNA (a perfluoroalkyl carboxylic acid) and (again) PFUdA vis-à-vis N-EtFOSSA, presumably is response to the lower hydrophobicity of the C8 phase functionality. Interestingly, both C8 phases and the PAH phase had fewer overlapping peaks compared to the C18 phases, but in different parts of the elution order spectrum. This likely represents the greater contribution of pi-electron interaction with the PAH phase in contrast with more consistent hydrophobic interaction characteristic of the C18 phases. These variations are subtle rather than dramatic, but they offer insights into interactions between solid phase chemistry and PFAS compound class that could be useful for better separating adjacent compound pairs or shifting analytes away from mass spectral interferences.

Figure 5.
C18 and PAH summary

		L-PFBS	PFHxA	HFPO-DA	PFHpA	L-PFHxS	NaDONA	PFOA	PFNA	L-PFOS	9-CI-PF3ONS	PFDA	N-MeFOSAA	PFUdA	N-EtFOSAA	11CI-PF3OUdS	PFDoA	PFTfDA	PFTeDA
Gemini™	C18	5.28	7.37	8.23	9.53	10.02	10.09	11.41	13.12	13.12	13.58	14.28	15.03	15.35	15.36	16.05	16.34	17.23	18.11
Luna™Omega	Polar C18	4.15	6.18	7.10	8.31	8.38	8.47	10.16	11.43	11.43	12.34	12.58	13.30	14.04	14.05	14.38	14.59	15.47	16.29
Luna Omega	Ps C18	4.29	6.34	7.21	8.46	8.55	9.05	10.35	12.06	12.03	12.53	13.24	14.01	14.31	14.36	14.58	15.23	16.08	16.44
Kinetex	C18	3.36	6.38	7.38	9.44	10.04	10.07	12.03	13.48	13.52	14.4	15.1	15.41	16.17	16.15	16.5	17.15	18.06	18.48
Kinetex	XB-C18	3.27	5.30	6.18	7.56	8.10	8.13	9.54	11.31	11.34	12.24	12.51	13.28	13.58	14.02	14.31	14.56	15.47	16.31
Kinetex	Polar C18	3.05	4.59	5.49	7.18	7.33	7.37	9.16	10.54	10.57	11.52	12.17	12.51	13.25	13.27	14.03	14.25	15.18	16.05

		L-PFBS	PFHxA	HFPO-DA	PFHpA	NaDONA	L-PFHxS	PFOA	PFNA	L-PFOS	9-CI-PF3ONS	PFDA	N-MeFOSAA	N-EtFOSAA	PFUdA	11CI-PF3OUdS	PFDoA	PFTfDA	PFTeDA
Kinetex	PAH	1.24	2.07	2.60	3.99	4.27	4.31	5.81	7.59	7.76	8.61	9.16	9.73	10.32	10.52	11.14	11.69	12.75	13.71

↑
Elution Order Shifts from C18
↑
Elution Order Shifts from C18
Overlaps are Eliminated

Figure 6.
C8 summary

		L-PFBS	PFHxA	HFPO-DA	PFHpA	L-PFHxS	NaDONA	PFOA	L-PFOS	PFNA	9-CI-PF3ONS	PFDA	N-MeFOSAA	N-EtFOSAA	PFUdA	11CI-PF3OUdS	PFDoA	PFTfDA	PFTeDA
Luna	C8	5.41	7.51	8.38	9.51	10.04	10.15	11.33	12.47	12.49	13.28	13.53	14.12	14.40	14.46	15.13	15.33	16.14	16.48
Kinetex	C8	5.30	7.54	8.47	10.14	10.26	10.33	11.57	13.18	13.19	14.03	14.26	14.45	15.15	15.24	15.56	16.14	16.56	17.33

↑
Elution Order Shifts from C18
↑

2. Column Chemistry Considerations (continued)

		L-PFBS	PFHxA	HFPO-DA	PFHpA	NaDONA	L-PFHxS	PFNA	PFNA	L-PFOS	PFDA	N-MeFOSAA	N-EFOSAA	9-CI-PF3ONS	PFUdA	PFDoA	11CI-PF3OUdS	PFTrDA	PFTeDA	
Kinetex™	PAH	0.76	0.76	0.83	0.95	1.05	1.33	1.34	2.19	3.44	3.49	3.78	4.30	4.31	4.78	6.10	6.90	7.47	8.82	
		 Elution Order Shifts from "EPA 533 Similar"																		

The methanol:acetonitrile mobile phase also resulted in more overlapping pairs and changes in elution order with the C8 columns (Figure 11). The elution order was consistent between the two C8 columns using this method, but there were many shifts in elution order compared to the methanol-only eluent. Finally, the methanol:acetonitrile method and the methanol-only method showed similar but not identical elution orders in the Kinetex Biphenyl and Phenyl-Hexyl Columns. The elution order with Kinetex F5 was less comparable with Kinetex Biphenyl and Phenyl-Hexyl columns with the acetonitrile altered eluent than previously seen with the methanol-only eluent. However, with the acetonitrile altered eluent, Kinetex F5 was more similar in elution order to the C18 columns than to the phenyl stationary phases.

The 533 Acetonitrile Altered method also showed increased overlapping peaks in all phenyl and F5 stationary phases (Figure 12), although the shorter run times may have contributed significantly to these increases. All things considered, the methanol:acetonitrile data demonstrate that mobile phase polarity (in conjunction with stationary phase chemistry) has a great deal of influence over the sorption behavior of the different classes of PFAS compounds and could be a powerful tool with which to influence chromatographic behavior.

Figure 9.
C8 Summary

		PFHxA	L-PFBS	HFPO-DA	PFHpA	NaDONA	PFOA	L-PFHxS	PFNA	PFDA	N-MeFOSAA	L-PFOS	N-EFOSAA	9-CI-PF3ONS	PFUdA	PFDoA	11CI-PF3OUdS	PFTrDA	PFTeDA	
Luna™	C8	2.05	2.15	2.34	3.22	3.55	4.51	5.07	6.22	7.48	7.53	8.04	8.28	9.05	9.09	10.25	11.33	11.36	12.42	
Kinetex	C8	2.14	2.22	2.41	3.30	4.10	5.19	5.34	7.01	8.32	8.35	8.47	9.11	9.50	9.53	11.06	12.17	12.14	13.17	
		 Elution Order Shifts from "EPA 533 Similar"																		

Figure 10.
Phenyl Stationary Phase Summary

		PFHxA	L-PFBS	HFPO-DA	PFHpA	NaDONA	PFOA	L-PFHxS	PFNA	PFDA	L-PFOS	N-MeFOSAA	N-EFOSAA	9-CI-PF3ONS	PFUdA	PFDoA	11CI-PF3OUdS	PFTrDA	PFTeDA	
Gemini™	C18	2.42	2.55	2.97	3.88	4.45	5.55	5.47	7.25	8.89	9.08	9.10	9.82	10.24	10.43	11.85	12.96	13.15	14.40	
		Benchmark: C 18 "EPA 533 Acetonitrile Altered"																		
		PFHxA	L-PFBS	HFPO-DA	PFHpA	NaDONA	PFOA	L-PFHxS	PFNA	PFDA	L-PFOS	N-MeFOSAA	9-CI-PF3ONS	N-EFOSAA	PFUdA	PFDoA	11CI-PF3OUdS	PFTrDA	PFTeDA	
Kinetex	F5	1.27	1.32	1.45	1.93	2.29	3.12	3.26	4.55	5.92	6.05	6.54	6.84	7.03	7.22	8.44	9.24	9.63	10.76	
		 Elution Order Shifts from C18 Acetonitrile Altered																		
		PFHxA	L-PFBS	HFPO-DA	PFHpA	NaDONA	PFOA	L-PFHxS	PFNA	PFDA	N-MeFOSAA	L-PFOS	N-EFOSAA	PFUdA	9-CI-PF3ONS	PFDoA	PFTrDA	11CI-PF3OUdS	PFTeDA	
Kinetex	Phenyl-Hexyl	1.05	1.14	1.18	1.39	1.60	1.99	2.25	2.96	4.09	4.33	4.47	4.82	5.16	5.46	6.17	7.09	7.45	7.95	
Kinetex	Biphenyl	0.85	0.90	0.90	0.99	1.07	1.20	1.32	1.55	2.11	2.32	2.37	2.68	2.81	3.23	3.59	4.31	4.83	5.05	
		 Elution Order Shifts from C18 Acetonitrile Altered																		

2. Column Chemistry Considerations (continued)

Conclusions

The HPLC methodology in EPA methods 537, 537.1 and 533 are all based upon a C18 stationary phase and a methanol-water mobile phase. In this study we have shown that the use of alternative stationary phases of varying surface chemistry and eluents of varying polarity can significantly alter the sorption-elution characteristics of different classes of PFAS compounds. This orthogonal approach to PFAS HPLC chromatography should serve as a fruitful avenue for future method development. As analyte lists increase in size and complexity, a variety of HPLC column chemistries and eluent compositions will be needed to accommodate the wide range of PFAS related compounds that might be encountered such as polar acids, non-polar acids, esters, amides, sulfonamides, and telomere length, all of which can be complicated with branched vs. linear isomers.

The work presented here is merely illustrative and should be considered a starting point for column chemistry and mobile phase considerations for PFAS HPLC methodology. Even though the demonstration sample contained a nice mix of PFAS compounds with varied functional groups, there are certainly many more compounds in the 9000-strong (and growing) PFAS inventory that will challenge LC-MS methodology. National and state PFAS analyte panels are constantly being updated and expanded. There is increasing emphasis on identifying and quantifying PFAS related isomers, unique functional groups and degradation products across a wide range of sample matrices. With regulated detection and quantitation limits being driven lower and lower, sensitivity is a significant issue. The choice of HPLC column chemistry will play a significant role in successfully meeting all these future challenges.

References

1. National Institute of Environmental Health Sciences. NIEHS Perfluorinated Chemicals (PFCs) fact sheet, 2012. https://www.atsdr.cdc.gov/sites/default/files/documents/perfluorinated_chemicals_508.pdf (accessed Dec 1, 2019).
2. United States Environmental Protection Agency. Per- and Polyfluoroalkyl Substances (PFAS). <https://www.epa.gov/pfas> (accessed Dec 1, 2019).
3. Agency for Toxic Substances and Disease Registry (ATSDR). An Overview of Perfluoroalkyl and Polyfluoroalkyl Substances and Interim Guidance for Clinicians Responding to Patient Exposure Concerns, 2019. https://www.atsdr.cdc.gov/pfas/docs/ATSDR_PFAS_ClinicalGuidance_12202019.pdf (accessed Apr 6, 2020).
4. PFAS List of PFAS Substances (Version 2). CompTox Chemicals Dashboard | PFASMASTER Chemicals (epa.gov) Accessed April 7, June 9, 2001.

Product Guide for PFAS Analysis

Phenomenex PFAS Products Referenced or Applicable in Official Methods

Regulatory Method	Product	Part Number
USEPA 537.1: Determination of Selected Per- and Polyfluorinated Alkyl Substances in Drinking Water by Solid Phase Extraction and Liquid Chromatography/ Tandem Mass Spectrometry (LC/MS/MS) (5)	PFAS CRM EPA 537.1 mix 1mL 2µg/mL in methanol	AL0-101839
	PFAS CRM EPA 533 + 537.1 mix 1mL 2µg/mL in methanol	AL0-101840
	Strata™ SDB-L 500 mg/6 mL	8B-S014-HCH
	Gemini™ 3 µm C18, 50 x 3 mm or	00B-4439-B0
USEPA Method 533: Determination of Per- and Polyfluoroalkyl Substances in Drinking Water by Isotope Dilution, Anion Exchange Solid Phase Extraction and LC-MS/MS. (1)	Luna™ Omega 1.6 µm PS C18 100 x 2.1 mm	00D-4752-AN
	PFAS CRM EPA 533 mix 1mL 2µg/mL in methanol	AL0-101838
	PFAS CRM EPA 533 + 537.1 mix 1mL 2µg/mL in methanol	AL0-101840
	Strata-X-AW 500 mg/6 mL	8B-S038-HCH
US Food and Drug Administration: Determination of 16 Perfluoroalkyl and Polyfluoroalkyl Substances(PFAS) in Food using Liquid Chromatography-Tandem Mass Spectrometry (LC-MS/MS). (2)	Gemini 3 µm C18 50 x 2 mm or	00B-4758-Y0
	Luna Omega 1.6 µm PS C18 100 x 2.1 mm	00D-4752-AN
	Strata-XL-AW 200 mg/3 mL	8B-S051-FBJ
US Department of Agriculture: Screening, Determination and Confirmation of PFAS by UPLC-MS-MS (3)	Luna C8(2) 3 µm 50 x 2 mm	00B-4248-B0
	Strata PFAS (WAX/GCB) 200 mg/50 mg/6 mL, 30/box 500 mg/50 mg/6 mL, 30/box	CS0-9207 CS0-9208
US Department of Defense: Quality Systems Manual (QSM) for Environmental Laboratories (4)	Gemini 3 µm C18 50 x 2 mm	00B-4439-B0

References

- [Method 537.1: Determination of Selected Per- and Polyfluorinated Alkyl Substances in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry \(LC/MS/MS\) | Science Inventory | US EPA](#)
- [Method 533: Determination of Per- and Polyfluoroalkyl Substances in Drinking Water by Isotope Dilution Anion Exchange Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry | Methods Approved to Analyze Drinking Water Samples to Ensure Compliance with Regulations | US EPA](#)
- [Determination of 16 Perfluoroalkyl and Polyfluoroalkyl Substances in Food using Liquid Chromatography-Tandem Mass Spectrometry \(fda.gov\)](#)
- [Screening, Determination and Confirmation of PFAS by UPLC-MS-MS \(usda.gov\)](#)
- <https://denix.osd.mil/edqw/documents/manuals/qsm-version-5-3-final/>

Recommended HPLC Products for Routine PFAS Analysis

Description and Function	Product	Part Number
Analytical Column (UHPLC)	Kinetex™ 5 µm EVO C18 100 x 2.1 mm	00D-4633-AN
	Luna Omega C18 1.6 µm 50 x 2.1	00B-4752-AN
	Gemini 3 µm C18 50 x 3 mm	00B-4439-Y0
Analytical Column	Gemini 3 µm C18 50 x 3 mm	00B-4439-Y0
Analytical Column (> 100 µL injection)	Gemini 3 µm C18 100 x 3 mm	00D-4439-Y0
Analytical Column (improved, low wt acids)	Luna Omega 3 µm PS C18 50 x 3 mm	00B-4758-Y0
Delay Column	Kinetex 5 µm EVO C18, 50 x 2.1 mm	00B-4633-AN 00A-4252-Y0
SecurityGuard	Luna Omega PS C18 4 x 3.0/10 pack for ID: 3.2-8.0 mm 4 x 2.0/10 pack for ID: 2.0-3.0 mm	AJ0-7606 AJ0-7605

Product Guide for PFAS Analysis (continued)

Recommended SPE Products

Description and Function	Product	Part Number
SPE Cartridge for EPA 537.1	Strata™ SDB-L 500 mg/6 mL, 30/box	8B-S014-HCH
SPE Cartridge for EPA 533	Strata-X-AW 33um Polymeric Weak Anion, 500 mg/6 mL, 30/box	8B-S038-HCH
SPE Cartridge (Rev. Phase, High Perf.)	Strata-XL 500 mg/6 mL, 30/box	8B-S043-HCH
SPE Stacked Cartridge (DOD QSM 5.3)	Strata PFAS (WAX/GCB) 200 mg/50 mg/6 mL, 30/box	CS0-9207
SPE Stacked Cartridge (DOD QSM 5.3)	Strata PFAS (WAX/GCB) 500 mg/50 mg/6 mL, 30/box	CS0-9208
SPE Cartridge (WAX for DOD QSM 5.3)	Strata-XL-AW 500 mg/6 mL, 30/box	8B-S051-HCH
GCB** Cartridge (GCB for DOD QSM 5.3)	Strata GCB 250 mg/6 mL, 30/box	8B-S528-FCH
SPE Cartridge (WAX* for FDA Method)	Strata-XL-AW 100 µm 200 mg/3 mL, 50/box	8B-S051-FBJ

(*WAX = Weak Anion Exchange)
(**GCB = Graphitized Carbon Black)

Recommended QuEChERS Products

Description and Function	Product	Part Number
QuEChERS Extraction (Soil/Sediment)	roQ QuEChERS Extraction Kit	KS0-8911
QuEChERS dSPE (Soil/Sediment)	roQ QuEChERS dSPE Kit, 15 mL	KS0-9516
QuEChERS Extraction (Dairy/Eggs/Fish)	roQ QuEChERS Extraction Kit	KS0-8910
QuEChERS dSPE (Dairy/Eggs/Fish)	roQ QuEChERS dSPE Kit	KS0-9511




Recommended Accessories

Description and Function	Product	Part Number
SPE Sample Reservoir	75 mL Sample Reservoir	H0-7005
Large Volume SPE	Adaptor Cap for 12,20, 60 mL SPE Tubes	AH0-7379
Autosampler Vials	Polypropylene, 300 µm + PE Starburst Cap	AR0-9995-12-C
Polypropylene Vials	Vial 9mm Screw Thd PP 2 mL, 1000 Pk	AR0-89C7-13
Vial Caps	Cap 9mm Solid Top Black Unlined	8B-S528-FCH
PEEK Capillary Tubing	Capillary Tubing Kit, Various Sizes	AT0-1964
PEEK Tubing Cutter	Cutter for PEEK Capillary Tubing	AT0-1110

Strata™ Solid Phase Extraction (SPE)

Strata-X

Ordering Information

Format	Sorbent Mass	Part Number	Unit
Tube			
	30 mg	8B-S100-TAK**	1 mL (100/box)
	30 mg	8B-S100-TBJ	3 mL (50/box)
	60 mg	8B-S100-UBJ**	3 mL (50/box)
	100 mg	8B-S100-EBJ	3 mL (50/box)
	100 mg	8B-S100-ECH	6 mL (30/box)
	200 mg	8B-S100-FBJ	3 mL (50/box)
	200 mg	8B-S100-FCH	6 mL (30/box)
	500 mg	8B-S100-HBJ	3 mL (50/box)
	500 mg	8B-S100-HCH	6 mL (30/box)
Giga™ Tube			
	500 mg	8B-S100-HDG	12 mL (20/box)
	1 g	8B-S100-JDG	12 mL (20/box)
	1 g	8B-S100-JEG	20 mL (20/box)
	2 g	8B-S100-KEG	20 mL (20/box)
	5 g	8B-S100-LFF	60 mL (16/box)
Teflon® Tube			
	200 mg	8B-S100-FBJ-T	3 mL (50/box)
	200 mg	8B-S100-FDG-T	12 mL (20/box)



On-line Extraction Cartridge

Description	Part Number	Unit/Box
Strata-X on-line extraction cartridge, 20 x 2.0 mm	00M-S033-B0-CB	ea
Cartridge holder, 20 mm	CHO-5845	ea

**Tab-less tubes available. Contact Phenomenex for details.

Strata-XL

Ordering Information

Format	Sorbent Mass	Part Number	Unit
Tube			
	30 mg	8B-S043-TAK	1 mL (100/box)
	60 mg	8B-S043-UBJ	3 mL (50/box)
	100 mg	8B-S043-EBJ	3 mL (50/box)
	200 mg	8B-S043-FBJ	3 mL (50/box)
	200 mg	8B-S043-FCH	6 mL (30/box)
	500 mg	8B-S043-HCH	6 mL (30/box)
Giga Tube			
	2 g	8B-S043-KDG	12 mL (20/box)
	2 g	8B-S043-KEG	20 mL (20/box)
	5 g	8B-S043-LEG	20 mL (20/box)
	5 g	8B-S043-LFF	60 mL (16/box)
	10 g	8B-S043-MFF	60 mL (16/box)
	30 mg	8E-S043-TGB	2 Plates/Box

* To control flow rate with Strata-XL, use a stopcock ([AHO-6048](#)) when processing samples with a vacuum manifold.

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A2LA Cert No. 2427.01



Gemini™ pH Flexible LC Columns

Ordering Information

3µm Microbore, Minibore and MidBore™ Columns (mm)								SecurityGuard™ Cartridges (mm)		
Phases	50 x 1.0	20 x 2.0	30 x 2.0	50 x 2.0	100 x 2.0	150 x 2.0	50 x 3.0	100 x 3.0	150 x 3.0	4 x 2.0* /10pk
C18	00B-4439-A0	00M-4439-B0	00A-4439-B0	00B-4439-B0	00D-4439-B0	00F-4439-B0	00B-4439-Y0	00D-4439-Y0	00F-4439-Y0	AJ0-7596

for ID: 2.0-3.0 mm

3µm Analytical Columns (mm)					SecurityGuard™ Cartridges (mm)	
Phases	30 x 4.6	50 x 4.6	100 x 4.6	150 x 4.6	250 x 4.6	4 x 3.0* /10pk
C18	00A-4439-E0	00B-4439-E0	00D-4439-E0	00F-4439-E0	00G-4439-E0	AJ0-7597

for ID: 3.2-8.0 mm



Kinetex™ Core-Shell LC Columns

Ordering Information

2.6µm Micro LC Columns (mm)						
Phases	30 x 0.3	50 x 0.3	100 x 0.3	150 x 0.3	50 x 0.5	150 x 0.5
EVO C18	—	00B-4725-AC	—	00F-4725-AC	00B-4725-AF	—

2.6µm MercuryMS™ LC-MS Cartridges (mm)		
Phases	20 x 2.0	20 x 4.0
Biphenyl	00M-4622-B0-CE	00M-4622-D0-CE

MercuryMS Cartridge Holders		
Part No.	Description	Unit
CHO-7188	Direct-Connect Cartridge Holder, 20 mm	ea
CHO-5845	Standard Cartridge Holder, 20 mm	ea

2.6µm Minibore Columns (mm)						SecurityGuard ULTRA Cartridges†
Phases	30 x 2.1	50 x 2.1	75 x 2.1	100 x 2.1	150 x 2.1	3/pk
EVO C18	00A-4725-AN	00B-4725-AN	—	00D-4725-AN	00F-4725-AN	AJ0-9298

for 2.1 mm ID

2.6µm MidBore™ Columns (mm)						SecurityGuard ULTRA Cartridges†
Phases	30 x 3.0	50 x 3.0	75 x 3.0	100 x 3.0	150 x 3.0	3/pk
EVO C18	00A-4725-Y0	00B-4725-Y0	—	00D-4725-Y0	00F-4725-Y0	AJ0-9297

for 3.0 mm ID

Ordering Information

5µm Minibore Columns (mm)					SecurityGuard™ ULTRA Cartridges‡
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk
EVO C18	00A-4633-AN	00B-4633-AN	00D-4633-AN	00F-4633-AN	AJ0-9298

for 2.1 mm ID

5µm MidBore™ Columns (mm)					SecurityGuard™ ULTRA Cartridges‡
Phases	30 x 3.0	50 x 3.0	100 x 3.0	150 x 3.0	3/pk
EVO C18	00A-4633-Y0	00B-4633-Y0	00D-4633-Y0	00F-4633-Y0	AJ0-9297

for 3.0 mm ID

5µm Analytical Columns (mm)					SecurityGuard™ ULTRA Cartridges‡
Phases	50 x 4.6	100 x 4.6	150 x 4.6	250 x 4.6	3/pk
EVO C18	00B-4633-E0	00D-4633-E0	00F-4633-E0	00G-4633-E0	AJ0-9296

for 4.6 mm ID



†SecurityGuard ULTRA Cartridges require holder, Part No.: [AJ0-9000](#)
 ‡SemiPrep SecurityGuard Cartridges require holder, Part No.: [AJ0-9281](#)
 ‡PREP SecurityGuard Cartridges require holder, Part No.: [AJ0-8223](#)

Luna™ One of The World's Leading LC Columns



Luna C18

Ordering Information

5 µm MidBore and Analytical Columns (mm)								SecurityGuard™ Cartridges (mm)	
Phases	30 x 3.0	50 x 3.0	150 x 3.0	250 x 3.0	30 x 4.6	50 x 4.6	75 x 4.6	4 x 2.0*	4 x 3.0*
C18(2)	00A-4252-Y0	00B-4252-Y0	00F-4252-Y0	00G-4252-Y0	00A-4252-E0	00B-4252-E0	00C-4252-E0	AJ0-4286 /10pk	AJ0-4287 /10pk
								for ID: 2.0-3.0 mm	3.2-8.0 mm

5 µm Analytical and Semi-Prep Columns (mm)					SecurityGuard™ Cartridges (mm)	
Phases	100 x 4.6	150 x 4.6	250 x 4.6	250 x 10	4 x 3.0*	10 x 10 ²
C18(2)	00D-4252-E0	00F-4252-E0	00G-4252-E0	00G-4252-N0	AJ0-4287 /10pk	AJ0-7221 /3pk
					for ID: 3.2-8.0 mm	9-16 mm

*SecurityGuard™ Analytical Cartridges require holder, Part No.: [KJ0-4282](#)
 *SemiPrep SecurityGuard™ Cartridges require holder, Part No.: [AJ0-9281](#)

Luna Omega PS C18 and Luna C18

Ordering Information

1.6 µm Microbore Columns (mm)			
Phases	50 x 1.0	100 x 1.0	150 x 1.0
PS C18	—	00D-4752-A0	—
C18	00B-4742-A0	00D-4742-A0	00F-4742-A0

1.6 µm Minibore Columns (mm)				SecurityGuard™ ULTRA Cartridges [†]	
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk
PS C18	00A-4752-AN	00B-4752-AN	00D-4752-AN	00F-4752-AN	AJ0-9508
C18	00A-4742-AN	00B-4742-AN	00D-4742-AN	00F-4742-AN	AJ0-9502
	for 2.1 mm ID				

Perfluoroalkyl Substances (PFAS) Testing Guide

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