TN-1285

Enhancing Sensitivity and Peak Capacity for Protein Digest using Micro-LC and the Power of 2.6 µm Kinetex[®] Core-Shell Columns

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Introduction

Microflow LC uses columns with smaller inner diameters and a reduced flow rate to achieve an improvement in ionization efficiency for LC-MS and as a result, greater sensitivity. In this tech note we show that coupling the ultra-high efficiency provided by the Kinetex core-shell 2.6 µm columns with these narrow column ID's can drive efficiency and peak capacity to improve what is seen for sub-2 µm fully porous UHPLC columns. High efficiency (narrower peaks) provides higher peak capacity, and this is something regarded as essential when looking at protein digest samples which are complex in nature and require detailed peak identification and profiling. When studying proteins, using a bottom-up proteomics approach, high efficiency reversed phase offers a significant advantage over other techniques by coupling high resolving power with mass spectrometry. Performance is measured by assessing peak capacity and this is defined as the number of peaks that can be separated or dispersed into the gradient time frame with a defined resolution.

The key to high efficiency with core-shell particles comes from their unique particle morphology and nature which minimizes efficiency losses and offers an increase in mass transfer rates through decreasing the effects of diffusion.

In this technote we compare the performance of the Kinetex $2.6\,\mu\text{m}$ core-shell packed into $0.3\,\text{mm}$ ID columns to a $1.7\,\mu\text{m}$ fully porous UHPLC alternative and demonstrate the average peak widths and peak capacities seen between a $2.6\,\mu\text{m}$ core-shell material are comparable to a sub- $2\,\mu\text{m}$ fully porous whilst operating at a significantly lower backpressure. Furthermore, the reduction in backpressure can facilitate the use of longer columns, something which has the potential to be extremely beneficial when the goal is maximising peak capacity.

Figure 1.

Comparison of average peak width and peak capacity for BSA tryptic digest

Column	Kinetex 2.6 µm XB-C18	Waters CSH M/Z Peptide 1.7 µm
Average Peak Width	0.184	0.191
Peak Capacity	54	45
Pressure (psi)	1900	4000

Results and Discussion

We compared the peak capacity and average peak width of the $2.6 \,\mu\text{m}$ Kinetex column to the $1.7 \,\mu\text{m}$ Waters[®] CSH and when using the 50 mm column length we found the average peak width to be lower and total peak capacity higher with the core-shell material (**Figure 1**). Peak capacity was calculated with the equation:

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PC = (T/PW)+1

where PC is the peak capacity, T is the retention time of the last eluting peak, and PW is the average peak width. **Figures 2** and **3** demonstrate an improved peak shape for many of the later eluting peptides when working with the Kinetex core-shell material leading to this increase in peak capacity. The lower backpressure observed when a 2.6 µm column is used allows additional column length to be investigated, unfortunately as a result of pressure in excess of 10,000 psi when using the 1.7 µm column, we were unable to obtain comparative data for 150 mm length using the Waters 1.7 µm nanoEase[™] M/Z Peptide CSH C18 column. When using the longer Kinetex column the gradient was adjusted accordingly to maintain a comparable gradient slope.

The 150 mm Kinetex $2.6 \,\mu$ m XB-C18 column allowed us to increase peak capacity from 54 to 105 when comparing it with the 50 mm alternative packed with the same media. There was a slight increase in peak width with increasing column length, however the overall increase in plate count because of a longer column affords significantly greater resolution across the gradient (**Figure 4**). Under these running conditions the pressure observed with the 150 x $0.3 \,\text{mm} 2.6 \,\mu$ m (4000 psi) column was comparable to the 50 x $0.3 \,\text{mm} 1.7 \,\mu$ m fully porous alternative (4000 psi), further highlighting the design space created when working with high efficiency core-shell particles.



Roxana Eggleston-Rangel, Application Scientist

Roxana likes to spend time with her dog and family. She has a German Shepherd mix named Cobi who was named after the official mascot of the 1992 Summer Olympics and not the basketball player. Besides the lab, you might find Roxana in old episodes of the Ghost Whisperer, MAD TV, The L word and others as she used to be a TV extra during her school years.



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Figure 2.

Kinetex® 2.6 µm XB-C18 50 x 0.3 mm, BSA tryptic digest



Figure 3. Waters[®] 1.7 μm nanoEase[™] M/Z Peptide CSH C18 column 50 x 0.3 mm, BSA tryptic digest





Temperature: 30 °C LC System: nanoLC[™] (SCIEX[®]) Detection: nanoESI Detector: 6500 QTRAP[®] (SCIEX) Injection Volume: 1 μL

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Kinetex[®] 2.6 µm XB-C18 150 x 0.3 mm, BSA tryptic digest



Conclusions

Kinetex columns packed with $2.6\,\mu m$ core-shell particles offer comparable efficiency and lower backpressure to fully porous $1.7\,\mu m$ columns with the same column dimension. The lower backpressure produced from a larger particle can be utilized to

allow longer columns to be used at flow rates which in many cases would be unattainable on sub-2 μm UHPLC columns due to pressure restrictions.



Ordering Information

2.6µm Micro LC Columns (mm)

Phases	30 x 0.3	50 x 0.3	100 x 0.3	150 x 0.3	50 x 0.5	150 x 0.5
XB-C18	00A-4496-AC	00B-4496-AC	00D-4496-AC	00F-4496-AC	00B-4496-AF	00F-4496-AF
Biphenyl		00B-4622-AC		00F-4622-AC	00B-4622-AF	_
C18	00A-4462-AC	00B-4462-AC		00F-4462-AC	00B-4462-AF	—
EV0 C18		00B-4725-AC		00F-4725-AC	00B-4725-AF	_
F5		00B-4723-AC	00D-4723-AC	00F-4723-AC	00B-4723-AF	_



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