

Wastewater, Sediment, and Soil

# Perfluoroalkyl Substances **(PFAS)** Testing Guide

# Wastewater, Sediment, and Soil

*As the PFAS story continues it is becoming more widely recognized that drinking water is not the only environmental media of concern. From its primary sources in fire suppression foams, industrial discharges and consumer products, PFAS is also widely found to occur in soils, sediments, surface water, groundwater and wastewater discharges, illustrating the widespread dispersion and persistence of this unique class of compounds. These discoveries have required the development and application of more advanced sample preparation, chromatography and mass spectrometry techniques to overcome the challenges of matrix and spectral interferences. In this section, two recent applications have been selected to illustrate the analytical challenges of these more difficult matrices.*



# 1. SPE for DOD QSM 5.3

## Per- and Polyfluoroalkyl Substances (PFAS) Extraction by LC-MS/MS Using Strata PFAS for a Stacked Solid Phase Extraction (SPE) Solution

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### Overview

PFAS are a class of highly stable synthetic organic compounds used in a wide variety of industrial and commercial applications. They are also highly stable in the environment and strongly bio-accumulate. As a result, they have become ubiquitous throughout the global environment and are often referred to in popular media as "Forever Chemicals". Consequently, PFAS levels need to be tested in drinking water and more recently methods have been developed to measure PFAS in other environmental matrices that require more complex clean-up solutions, such as wastewater, soils and sediments.



The United States Department of Defense (DOD) is dealing with very extensive PFAS contamination owing to the widespread use of PFAS based Aqueous Film Forming Foam (AFFF) used as fire suppression foams at many military installations. As a result, DOD has developed its own PFAS analytical guidelines to deal with the unique environmental monitoring and clean-up challenges found on their installations. These guidelines, contained within the DOD QSM 5.1/5.3 documentation (Department of Defense QSM (osd.mil), feature a unique sample clean-up and concentration approach not found in EPA Methods which are designed only for drinking water application.

DOD QSM 5.1/5.3 specifies the use of a polymeric weak anion exchange (WAX) SPE sorbent in combination with graphitized carbon black (GCB) sorbent for the clean-up of solid samples, soils, biota, sediments, or non-drinking water samples. This can be performed by using two individual tubes of WAX and GCB sorbent that are applied sequentially or the use of dispersive SPE (dSPE) utilizing GCB following the WAX SPE tube extraction. Both methods add time to the clean-up procedure and present the opportunity for loss of analytes and introduction of imprecision. In this communication we describe a significant improvement to the guidelines, Strata PFAS SPE, wherein the two sorbents are contained within a single tube, offering the opportunity for decreased sample processing time and increased accuracy and precision. When comparing recoveries for a small subset of analytes for a WAX SPE and dSPE GCB method vs Strata™ PFAS, the recovery is greatly improved for Strata PFAS (**Table 1**).

Strata PFAS is a stacked single cartridge solution with polymeric WAX and GCB sorbents that functions as a traditional Solid Phase Extraction (SPE) cartridge with a built in polishing step to meet the aforementioned DOD guidelines. This SPE product increases lab productivity and reduces the need for multiple extraction tubes when compared to a traditional two tube method.

**Table 2** presents typical analyte recovery data from a routine Laboratory Control Sample (LCS) analyzed by a commercial testing laboratory highly experienced with the performance of DOD QSM 5.1/5.3. The LCS had been spiked with all 32 target analytes at 25 µg/L and was analyzed with a batch of field samples to demonstrate method performance and data acceptability. The recovery data show that all 32 analytes were well within method recovery limits with an average recovery of 98.8 % and a mean recovery of 99.0 %, thereby demonstrating acceptability of the use of Strata PFAS in the performance DOD QSM5.1/5.3.

The LCS sample was extracted with Strata PFAS under the conditions shown below and analyzed on a LC-MS/MS system using a Gemini™ 3 µm C18 HPLC column under the conditions described below.

### SPE Conditions

**Cartridge:** Strata PFAS (200 mg WAX/50 mg GCB/ 6 mL)

**Part No.:** [CSO-9207](#)

**Condition 1:** 4 mL 0.3% Ammonium hydroxide

**Condition 2:** 4 mL Methanol

**Equilibrate:** 5 mL Water

**Load:** Add sample at 4 mL/min

**Wash:** 2x 4 mL Water

**Elute:** 2x 4 mL 0.3% Ammonium hydroxide in Methanol

**Evaporate:** To dryness and reconstitute to 1 mL with

Methanol/Water (96:4)

### LC-MS/MS Parameters

**Column:** Gemini 3 µm C18

**Dimensions:** 50 x 2.0 mm

**Part No.:** [00B-4439-B0](#)

**Mobile Phase:** A: 20 mM Ammonium acetate in Water

B: Methanol

Gradient: Time (min)	%B
0	5
0.1	55
4.5	99
8.0	99
8.5	5

**Flow Rate:** 0.6 mL/min

**Delay column:** Luna™ 5 µm C18(2) 30 x 3.0 mm ([00A-4252-Y0](#)) installed between the autosampler and mobile phase pump mixer

**Injection Volume:** 10 µL

### Mass Spec Parameters

**Mass Spec Detector:** SCIEX® Triple Quad™ 4500

**Ion Source Parameters:** Samples were ionized using electrospray in negative ion-mode

Parameter	Value
CAD	9
CUR	30
GS1	40
GS2	60
IS Voltage	-4500
TEM	450

### MRM Transitions for HFPO-DA

Compound	Q1	Q3	RT	DP	CE
HFPO-DA (Quant)	329	185	3.7	-30	-32
HFPO-DA (Qual)	329	169	3.7	-30	-18
13C3 -HFPO-DA	332	185	3.7	-30	-32

# 1. SPE for DOD QSM 5.3 (continued)

**Table 1.**

**Recovery Comparisons of WAX SPE and dSPE using GCB vs Strata PFAS Single Cartridge Method**

Analyte	WAX SPE + dSPE GCB % Recovery	Strata PFAS Stacked Cartridge % Recovery
13C2-PFDoDA	77.0	84.5
13C2-PFTeDA	62.0	84.0
PFDoDA	38.0	78.3
PFHxDA	63.0	89.3

**Table 2.**

**Recovery of QSM 5.3 Target Analytes from a Laboratory Control Sample Using Strata PFAS SPE (WAX/GCB)**

Analyte	Actual Concentration	Sample Result	% Recovery	Method Limits	Pass/Fail
PFBA	25.600	22.640	88	84-135	Pass
PPPeA	25.600	22.157	87	75-138	Pass
PFBS	22.640	22.300	99	81-133	Pass
4:2-FTS	23.920	22.078	92	64-134	Pass
PFHxA	25.600	24.644	96	80-137	Pass
PFPes	24.000	21.699	90	82-132	Pass
HFPoDA	25.600	26.336	103	0-130	Pass
PFHpA	25.600	27.018	106	80-140	Pass
PFHpA	25.600	27.018	106	80-140	Pass
PFHxS	24.200	24.713	102	71-131	Pass
DONA	24.120	26.083	108	70-130	Pass
6:2-FTS	24.280	24.217	100	51-155	Pass
PFHpS	24.360	23.015	94	80-129	Pass
PFOA	25.600	25.043	98	83-138	Pass
PFOS	24.480	22.492	92	54-139	Pass
PFNA	25.600	25.872	101	73-140	Pass
9Cl-PF3ONS	23.840	21.863	92	70-130	Pass
PFNS	24.560	21.993	90	71-121	Pass
PFNS	24.560	21.993	90	71-121	Pass
PFDA	25.600	25.047	98	78-137	Pass
8:2-FTS	24.520	22.231	91	62-133	Pass
PFOSA	25.600	25.714	100	73-121	Pass
NMEFOSAA	25.600	30.906	121	53-136	Pass
PFDS	24.640	22.873	93	69-124	Pass
PFUnDA	25.600	26.353	103	70-134	Pass
NEtFOSAA	25.600	28.765	112	59-145	Pass
11Cl-PF30UDS	24.120	22.625	94	70-130	Pass
PFDoDA	25.600	27.710	108	75-139	Pass
10:2-FTS	24.680	26.626	108	50-124	Pass
PFDoS	24.800	21.509	87	39-121	Pass
PFTrDA	25.600	25.814	101	67-144	Pass
PFTeDA	25.600	25.446	99	79-134	Pass
PFDoDA	25.600	27.373	107	10-124	Pass

Recovery Range: 87% - 116%

Average Recovery: 98.8%

Mean Recovery: 99.0%

## 2. Determination of PFAS in Sediments

### Determination of Perfluoroalkyl Substances (PFAS) in Sediments by QuEChERS Extraction and HPLC-MS/MS

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#### Introduction

Perfluoroalkyl substances (PFAS) are a class of highly stable synthetic organic compounds used in a wide variety of industrial and commercial applications including surface treatment for textiles, packaging materials, and non-stick cookware. PFAS are characterized by a hydrophobic fully fluorinated alkyl chain and a hydrophilic functional group. They are persistent in the environment due to the exceptional stability of the C-F bond. Once released in the aquatic environment, these chemicals will partition between the water phase and the sediment.



Currently, there are no federal regulatory limits controlling the discharge of PFAS compounds into the environment. Looking forward, it is possible that at some point EPA may establish regulatory limits for the various PFAS compounds in drinking water, wastewater and solid waste. In anticipation of such future developments, it is prudent to develop robust analytical methods and begin to better understand the fate and transport of these compounds in both the solid and liquid environmental fractions.

There are several methods available for the extraction and analysis of PFAS in aqueous samples, including the EPA Methods 537.1 and 533 previously described in this Guide (5). However, very few procedures are available for extracting these compounds in solid matrices such as sediments (1). Typical methods used are mechanical shaker and ultrasonic-assisted Solid-Liquid Extractions (SLE) 3, 4, 5). The extracts are then subjected to additional cleanup steps, usually by solid phase extraction, such as in the DoD QSM 5.3 approach previously described in this Guide. These are generally solvent-intensive and time-consuming processes. However, in 2003, an extraction procedure called QuEChERS (Quick-Easy-Cheap-Effective-Rugged-and-Safe) developed by researchers at the US Department of Agriculture was introduced (6). It was originally developed to extract pesticide residues in food matrices but has since found many other applications in the field of environmental analytical chemistry.

Our laboratory (LACSD) previously developed and validated a QuEChERS sediment extraction procedure for emerging contaminants including: pharmaceutical and personal care products, steroids, alkylphenol ethoxylates, and pyrethroid pesticides 7,8,9). We have successfully applied the same extraction method to determine perfluoroalkyl substances in marine and freshwater sediments.

#### Materials and Methods

##### Reagents/Chemicals

- QuEChERS Extraction – In a 50 mL plastic centrifuge tube combine 2.0 g of Anhydrous Magnesium Sulfate, and 1.5 g Sodium Acetate or use approximately 3.5 g of AOAC 2007.01 roQ™ extraction packet (part no. [AHO-9043](#))
- QuEChERS dSPE Clean-Up – roQ 15 mL dSPE Kit (part no. [KS0-8926](#))

#### Sample Preparation

##### QuEChERS Extraction Protocol

1. Weigh 2.0g of dried sediment into a polypropylene container and spike with isotopically-labeled internal standards. PPCPs, Steroids, and Pyrethroids can be extracted concurrently with this method by adding the appropriate internal standard and spiking solutions to the samples and QC's 7,8,9.
2. Add 10mL deionized water and vortex. Add 10mL acidified acetonitrile (1% acetic acid) to the slurry and vortex.
3. Add the extraction salts (1.5g Sodium Acetate and 2 g MgSO<sub>4</sub>) to the sample and vortex for 1 minute.
4. Centrifuge the samples for 5 minutes at 4000 rpm.
5. Place the samples in a rack and freeze at -20° for 30-60 minutes. This freezing step allows for easier extraction of the supernatant.
6. Transfer 8-9 mL of the acetonitrile supernatant into a roQ QuEChERS PSA/C18 dSPE clean-up tube (Part no. [KS0-8926](#)) and vortex for one minute.
7. Centrifuge the dSPE tubes for 10 minutes at 3000 rpm.
8. Place an aliquot of the extract in a HPLC vial and dilute 1:1 with deionized water. The sample is now ready for analysis.

#### HPLC-MS/MS Conditions

Column:	Gemini™ 3 µm C18
Dimensions:	100 x 3 mm
Part No.:	<a href="#">00D-4439-Y0</a>
Inline Filter:	Phenomenex Krudkatcher™ Ultra
Delay Column:	Luna™ 5 µm C18 (2) 30 x 2.0 mm
Part No.:	<a href="#">00A-4252-B0</a>
Mobile Phase:	A: 20 mM Ammonium acetate in water B: Methanol
Gradient:	Time (min) % B 0.0 10 1.5 65 8.0 95 8.1 99 12.0 99 12.5 10
Injection:	90 µL
Flow Rate:	0.6 mL/min
Temperature:	40 °C
Detector:	SCIEX® 5500 QTRAP®
Detection:	MS/MS ESI Negative (sMRM)
Analytes:	1. PFBA 2. PFPeA 3. PFBS 4. PFHxA 5. PPFS 6. PFHxS 7. PFHpA 8. PFHpS 9. PFOA

## 2. Determination of PFAS in Sediments (continued)

### Mass Spectrometer Parameters

**Table 1.**  
**MRM Transitions and Compound Dependent Parameters**

Compound Name	Q1	Q3	DP	CE
Perfluorobutanoic acid (PFBA)	213	169	-71	-14
Perfluoropentanoic acid (PFPeA)	263	219	-71	-12
Perfluorohexanoic acid (PFHxA)	313	269	-60	-14
Perfluoroheptanoic acid (PFHpA)	363	319	-62	-15
Perfluorooctanoic acid (PFOA)	413	369	-91	-12
Perfluorononanoic acid (PFNA)	463	419	-79	-15
Perfluorodecanoic acid (PFDA)	513	469	-83	-17
Perfluoroundecanoic acid (PFUdA)	563	519	-60	-17
Perfluorododecanoic acid (PFDoA)	613	569	-50	-21
Perfluorotridecanoic acid (PFTrDA)	663	619	-49	-18
Perfluorotetradecanoic acid (PFTeDA)	713	669	-63	-20
Perfluorobutanesulfonate (PFBS)	299	80	-94	-70
Perfluoropentantenesulfonate (PFPeS)	349	80	-96	-66
Perfluorohexanesulfonate (PFHxS)	399	80	-92	-75
Perfluoroheptanesulfonate (PFHpS)	449	80	-75	-84
Perfluoroctanesulfonate (PFOS)	499	80	-78	-96
Perfluorooctanesulfonamide (PFOSA)	498	78	-60	-84
Perfluorononanesulfonate (PFNS)	549	80	-87	-100
Perfluorodecanesulfonate (PFDS)	599	80	-55	-100
Perfluoro-n- <sup>13</sup> C4-butanoic acid (M4PFBA)	217	171.9	-71	-13
Perfluoro-n- <sup>13</sup> C5-pentanoic acid (M5PFPeA)	268	222.7	-71	-12
Perfluoro-n- <sup>13</sup> C5-hexanoic acid (M5PFHxA)	318	272.9	-60	-13
Perfluoro-n- <sup>13</sup> C4-heptanoic acid (M4PFHpA)	367	321.8	-62	-14
Perfluoro-n- <sup>13</sup> C8-octanoic acid (M8PFOA)	421	376	-91	-12
Perfluoro-n- <sup>13</sup> C9-nonanoic acid (M9PFNA)	472	427	-79	-17
Perfluoro-n- <sup>13</sup> C6-decanoic acid (M6PFDA)	519	474	-83	-21
Perfluoro-n- <sup>13</sup> C7-undecanoic acid (M7PFUdA)	570	525	-60	-17
Perfluoro-n- <sup>13</sup> C2-dodecanoic acid (M2PFDoA)	615	570	-50	-24
Perfluoro-n- <sup>13</sup> C2-tetradecanoic acid (M2PFTeDA)	715	670	-63	-25
Perfluoro- <sup>13</sup> C3-butanethiosulfonate (M3PFBS)	302	80	-94	-55
Perfluoro- <sup>13</sup> C3-hexanesulfonate (M3PFHxS)	402	80	-92	-85
Perfluoro- <sup>13</sup> C8-octanesulfonate (M8PFOS)	507	80	-78	-100

Note: DP = Declustering Potential

CE = Collision Energy

**Table 2.**  
**MS Source Parameters**

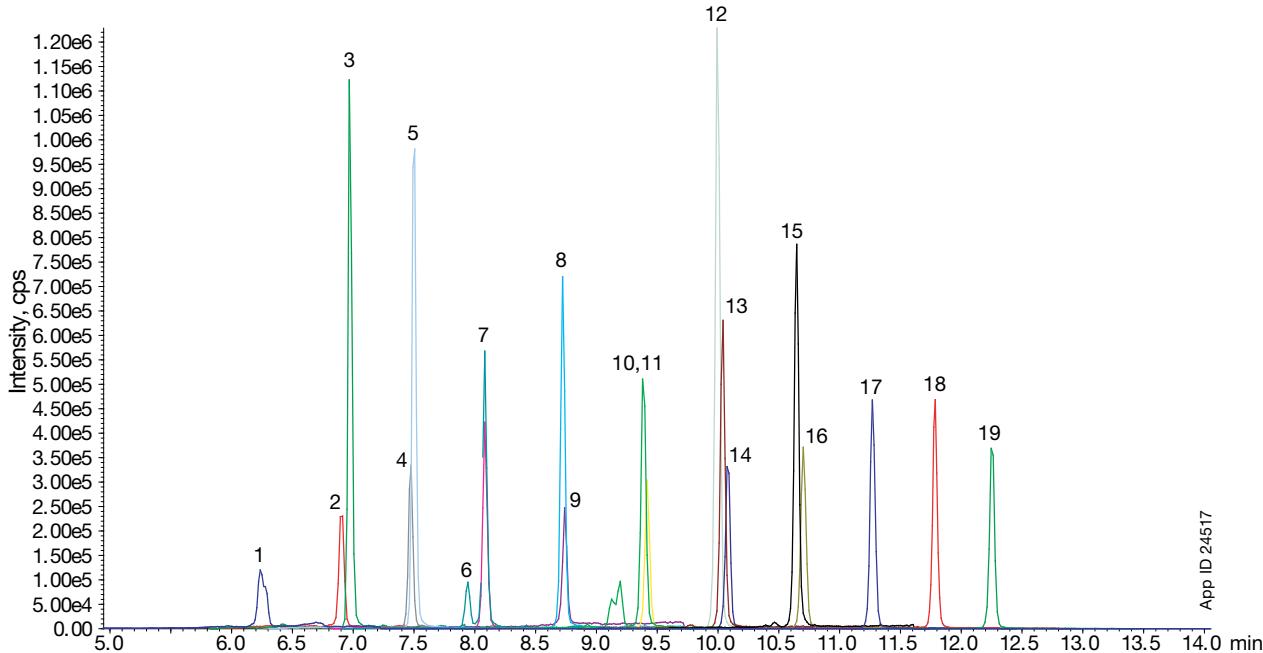
Source Parameters	Settings
Temperature	400 °C
Gas 1	50
Gas 2	50
Curtain Gas	35
Ionization Energies	-4500 V
Collision Gas	High

**Table 3.**  
**Method Performance Data for Sediments Spiked at 1 ng/g of the Target Analytes (n=4)**

Compound	Average % Recovery	% RSD
PFBA	91.7	0.76
PFPeA	86.3	6
PFHxA	89.4	1.2
PFHpA	93.1	2.9
PFOA	98.3	1.5
PFNA	93	1.6
PFDA	87.7	4.5
PFUdA	92.3	2.1
PFDoA	92.5	4.1
PFTrDA	88.2	2.1
PFTeDA	87.6	2.1
PFBS	86.3	2.1
PFPeS	96.2	3.2
PFHxS	81.3	5
PFHpS	92.3	2.6
PFOS	92.1	2.6
PFOSA	104.5	6.3
PFNS	89.8	6.8
PFDS	87.3	6.7

## 2. Determination of PFAS in Sediments (continued)

**Figure 1.**  
Extracted ion chromatogram of sediments spiked with 1.0ng/g of the target analytes



### Results and Discussion

QuEChERS is a vortex-assisted solid-liquid extraction procedure that uses acetonitrile, salts, and buffering agents for extraction, phase-separation, and pH adjustment respectively. Extracts are subsequently transferred to a dispersive solid phase extraction (dSPE) tube containing a drying agent ( $\text{MgSO}_4$ ) and SPE sorbents such as C18 or PSA for sample cleanup.

The modified QuEChERS method presented here is a simple, efficient, and cost-effective method for determining PFAS levels in sediments. Accuracy and precision were assessed using four replicates of sediments spiked with the target analytes. Average % recoveries are all within the 80–120 % range and % RSDs for all analytes are below 10 % (Table 3). Reporting limits were set at 0.05 ng/g dry weight based on a 2.0 g initial sample weight.

### References

1. Roberts S, et al. Quantitation of PFAS in Water Samples using LC/MS/MS: Large Volume Direct Injection and Solid Phase Extraction (2016) SCIEX Application Note Publication Number: RUO-MKT-02-4707-A
2. Berlizou-Barbier, A., et al., Multi-residue analysis of emerging pollutants in sediment using QuEChERS-based extraction followed by LC-MS/MS analysis. Analytical and Bioanalytical Chemistry (2014) 406:1259-1266
3. Jahnke A, et al. Trace analysis of per- and polyfluorinated alkyl substances in various matrices How do current methods perform? Journal of Chromatography A (2009) 1216: 410-4213. Estil S, et al, A. Rapid Extraction and Analysis of Steroids and Pyrethroids
4. Ahrens et. al. Partitioning behavior of per- and polyfluoroalkyl compounds between pore water and sediment in two sediment cores from Tokyo Bay, Japan. Environmental Science and Technology (2009) 43: 6969-6975
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6. Lehotay, S.J., Mastovska, K., Lightfield, A.R., Use of Buffering to Improve Results of Problematic Pesticides in a Fast and Easy Method for Residue Analysis of Fruits and Vegetables. Journal of Official Analytical Chemists International. 88.pp. 615-629.
7. Estil S, et al., A. Rapid Extraction and Analysis of PPCPs from Sediments by QuEChERS and LC/MS/MS (2016) Phenomenex Application Note: TN-0099 from Sediments by QuEChERS and LC/MS/MS (2016) Phenomenex Application Note: TN-0096 & TN-0098
8. Estil, S., et. al., A. Rapid Extraction of Steroids and Pyrethroids from Sediments by QuEChERS and LC/MS/MS (2016) Phenomenex Application Note: TN-0098
9. Estil S, et al, A. Rapid Extraction and Analysis of Pyrethroids from Sediments by QuEChERS and LC/MS/MS (2016) Phenomenex Application Note: TN-0098

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# Product Guide for PFAS Analysis

## Phenomenex PFAS Products Referenced or Applicable in Official Methods

Regulatory Method	Product	Part Number
USEPA 537.1: Determination of Selected Per-and Polyfluorinated Alkyl Substances in Drinking Water by Solid Phase Extraction and Liquid Chromatography/ Tandem Mass Spectrometry (LC/MS/MS) (5)	PFAS CRM EPA 537.1 mix 1mL 2µg/mL in methanol	A00-101839
USEPA Method 533: Determination of Per- and Polyfluoroalkyl Substances in Drinking Water by Isotope Dilution, Anion Exchange Solid Phase Extraction and LC-MS/MS. (1)	PFAS CRM EPA 533 + 537.1 mix 1mL 2µg/mL in methanol Strata™ SDB-L 500 mg/6 mL Gemini™ 3 µm C18, 50 x 3 mm or Luna™ Omega 1.6 µm PS C18 100 x 2.1 mm	A00-101840 8B-S014-HCH 00B-4439-B0 00D-4752-AN
US Food and Drug Administration: Determination of 16 Perfluoroalkyl and Polyfluoroalkyl Substances(PFAS) in Food using Liquid Chromatography-Tandem Mass Spectrometry (LC-MS/MS). (2)	PFAS CRM EPA 533 mix 1mL 2µg/mL in methanol	A00-101838
US Department of Agriculture: Screening, Determiation and Confirmation of PFAS by UPLC-MS-MS (3)	PFAS CRM EPA 533 + 537.1 mix 1mL 2µg/mL in methanol Strata-X-AW 500 mg/6 mL Gemini 3 µm C18 50 x 2 mm or Luna Omega 1.6 µm PS C18 100 x 2.1 mm	A00-101840 8B-S038-HCH 00B-4758-Y0 00D-4752-AN
US Department of Defense: Quality Systems Manual (QSM) for Environmental Laboratories (4)	Strata-XL-AW 200 mg/3 mL Luna C8(2) 3 µm 50 x 2 mm Strata PFAS (WAX/GCB) 200 mg/50 mg/6 mL, 30/box 500 mg/50 mg/6 mL, 30/box Gemini 3 µm C18 50 x 2 mm	8B-S051-FBJ 00B-4248-B0 CSO-9207 CSO-9208 00B-4439-B0

## References

- [Method 537.1: Determination of Selected Per- and Polyfluorinated Alkyl Substances in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry \(LC/MS/MS\) | Science Inventory | US EPA](#)
- [Method 533: Determination of Per- and Polyfluoroalkyl Substances in Drinking Water by Isotope Dilution Anion Exchange Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry | Methods Approved to Analyze Drinking Water Samples to Ensure Compliance with Regulations | US EPA](#)
- [Determination of 16 Perfluoroalkyl and Polyfluoroalkyl Substances in Food using Liquid Chromatography-Tandem Mass Spectrometry \(fda.gov\)](#)
- [Screening, Determination and Confirmation of PFAS by UPLC-MS-MS \(usda.gov\)](#)
- <https://denix.osd.mil/edqw/documents/manuals/qsm-version-5-3-final/>

## Recommended HPLC Products for Routine PFAS Analysis

Description and Function	Product	Part Number
Analytical Column (UHPLC)	Kinetex™ 5 µm EVO C18 100 x 2.1 mm Luna Omega C18 1.6 µm 50 x 2.1 Gemini 3 µm C18 50 x 3 mm	00D-4633-AN 00B-4752-AN 00B-4439-Y0
Analytical Column	Gemini 3 µm C18 50 x 3 mm	00B-4439-Y0
Analytical Column (> 100 µL injection)	Gemini 3 µm C18 100 x 3 mm	00D-4439-Y0
Analytical Column (improved. Imwt acids)	Luna Omega 3 µm PS C18 50 x 3 mm	00B-4758-Y0
Delay Column	Kinetex 5 µm EVO C18, 50 x 2.1 mm	00B-4633-AN 00A-4252-Y0
SecurityGuard	Luna Omega PS C18 4 x 3.0/10 pack for ID: 3.2-8.0 mm 4 x 2.0/10 pack for ID: 2.0-3.0 mm	AJ0-7606 AJ0-7605

# Product Guide for PFAS Analysis (continued)

## Recommended SPE Products

Description and Function	Product	Part Number
SPE Cartridge for EPA 537.1	Strata™ SDB-L 500 mg/6 mL, 30/box	<a href="#">8B-S014-HCH</a>
SPE Cartridge for EPA 533	Strata-X-AW 33um Polymeric Weak Anion, 500 mg/6 mL, 30/box	<a href="#">8B-S038-HCH</a>
SPE Cartridge (Rev. Phase, High Perf.)	Strata-XL 500 mg/6 mL, 30/box	<a href="#">8B-S043-HCH</a>
SPE Stacked Cartridge (DOD QSM 5.3)	Strata PFAS (WAX/GCB) 200 mg/50 mg/6 mL, 30/box	<a href="#">CS0-9207</a>
SPE Stacked Cartridge (DOD QSM 5.3)	Strata PFAS (WAX/GCB) 500 mg/50 mg/6 mL, 30/box	<a href="#">CS0-9208</a>
SPE Cartridge (WAX for DOD QSM 5.3)	Strata-XL-AW 500 mg/6 mL, 30/box	<a href="#">8B-S051-HCH</a>
GCB** Cartridge (GCB for DOD QSM 5.3)	Strata GCB 250 mg/6 mL, 30/box	<a href="#">8B-S528-FCH</a>
SPE Cartridge (WAX* for FDA Method)	Strata-XL-AW 100 µm 200 mg/3 mL, 50/box	<a href="#">8B-S051-FBJ</a>

(\*WAX = Weak Anion Exchange)

(\*\*GCB = Graphitized Carbon Black)

## Recommended QuEChERs Products

Description and Function	Product	Part Number
QuEChERs Extraction (Soil/Sediment)	roQ QuEChERs Extraction Kit	<a href="#">KS0-8911</a>
QuEChERs dSPE (Soil/Sediment)	roQ QuEChERs dSPE Kit, 15 mL	<a href="#">KS0-9516</a>
QuEChERs Extraction (Dairy/Eggs/Fish)	roQ QuEChERs Extraction Kit	<a href="#">KS0-8910</a>
QuEChERs dSPE (Dairy/Eggs/Fish)	roQ QuEChERs dSPE Kit	<a href="#">KS0-9511</a>

## Recommended Accessories

Description and Function	Product	Part Number
SPE Sample Reservoir	75 mL Sample Reservoir	<a href="#">H0-7005</a>
Large Volume SPE	Adaptor Cap for 12,20, 60 mL SPE Tubes	<a href="#">AHO-7379</a>
Autosampler Vials	Polypropylene, 300 µm + PE Starburst Cap	<a href="#">ARO-9995-12-C</a>
Polypropylene Vials	Vial 9mm Screw Thd PP 2 mL, 1000 Pk	<a href="#">ARO-89C7-13</a>
PEEK Capillary Tubing	Capillary Tubing Kit, Various Sizes	<a href="#">ATO-1964</a>
PEEK Tubing Cutter	Cutter for PEEK Capillary Tubing	<a href="#">ATO-1110</a>

# Strata™ Solid Phase Extraction (SPE)

## Strata-X

### Ordering Information

Format	Sorbent Mass	Part Number	Unit
<b>Tube</b>			
	30 mg	8B-S100-TAK**	1 mL (100/box)
	30 mg	8B-S100-TBJ	3 mL (50/box)
	60 mg	8B-S100-UBJ**	3 mL (50/box)
	100 mg	8B-S100-EBJ	3 mL (50/box)
	100 mg	8B-S100-ECH	6 mL (30/box)
	200 mg	8B-S100-FBJ	3 mL (50/box)
	200 mg	8B-S100-FCH	6 mL (30/box)
	500 mg	8B-S100-HBJ	3 mL (50/box)
	500 mg	8B-S100-HCH	6 mL (30/box)
<b>Giga™ Tube</b>			
	500 mg	8B-S100-HDG	12 mL (20/box)
	1 g	8B-S100-JDG	12 mL (20/box)
	1 g	8B-S100-JEG	20 mL (20/box)
	2 g	8B-S100-KEG	20 mL (20/box)
	5 g	8B-S100-LFF	60 mL (16/box)
<b>Teflon® Tube</b>			
	200 mg	8B-S100-FBJ-T	3 mL (50/box)
	200 mg	8B-S100-FDG-T	12 mL (20/box)

## Strata-XL

### Ordering Information

Format	Sorbent Mass	Part Number	Unit
<b>Tube</b>			
	30 mg	8B-S043-TAK	1 mL (100/box)
	60 mg	8B-S043-UBJ	3 mL (50/box)
	100 mg	8B-S043-EBJ	3 mL (50/box)
	200 mg	8B-S043-FBJ	3 mL (50/box)
	200 mg	8B-S043-FCH	6 mL (30/box)
	500 mg	8B-S043-HCH	6 mL (30/box)
<b>Giga Tube</b>			
	2 g	8B-S043-KDG	12 mL (20/box)
	2 g	8B-S043-KEG	20 mL (20/box)
	5 g	8B-S043-LEG	20 mL (20/box)
	5 g	8B-S043-LFF	60 mL (16/box)
	10 g	8B-S043-MFF	60 mL (16/box)
	30 mg	8E-S043-TGB	2 Plates/Box

\* To control flow rate with Strata-XL, use a stopcock ([AHO-6048](#)) when processing samples with a vacuum manifold.

### On-line Extraction Cartridge

Description	Part Number	Unit/Box
Strata-X on-line extraction cartridge, 20 x 2.0 mm	00M-S033-B0-CB	ea
Cartridge holder, 20 mm	CHO-5845	ea

\*\*Tab-less tubes available. Contact Phenomenex for details.

Currently offered by Phenomenex in USA and Canada only. Other regions coming soon.

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# Gemini™ pH Flexible LC Columns

## Ordering Information

3 µm Microbore, Minibore and MidBore™ Columns (mm)										SecurityGuard™ Cartridges (mm)	
Phases	50 x 1.0	20 x 2.0	30 x 2.0	50 x 2.0	100 x 2.0	150 x 2.0	50 x 3.0	100 x 3.0	150 x 3.0	4 x 2.0* /10pk	
C18	00B-4439-A0	00M-4439-B0	00A-4439-B0	00B-4439-B0	00D-4439-B0	00F-4439-B0	00B-4439-Y0	00D-4439-Y0	00F-4439-Y0	AJ0-7596	

for ID: 2.0-3.0 mm

3 µm Analytical Columns (mm)						SecurityGuard™ Cartridges (mm)	
Phases	30 x 4.6	50 x 4.6	100 x 4.6	150 x 4.6	250 x 4.6	4 x 3.0* /10pk	
C18	00A-4439-E0	00B-4439-E0	00D-4439-E0	00F-4439-E0	00G-4439-E0	AJ0-7597	

for ID: 3.2-8.0 mm



# Kinetex™ Core-Shell LC Columns

## Ordering Information

2.6 µm Micro LC Columns (mm)					
Phases	30 x 0.3	50 x 0.3	100 x 0.3	150 x 0.3	50 x 0.5
EVO C18	—	00B-4725-AC	—	00F-4725-AC	00B-4725-AF

2.6 µm MercuryMS™ LC-MS Cartridges (mm)		
Phases	20 x 2.0	20 x 4.0
Biphenyl	00M-4622-B0-CE	00M-4622-D0-CE

MercuryMS Cartridge Holders		
Part No.	Description	Unit
CHO-7188	Direct-Connect Cartridge Holder, 20 mm	ea
CHO-5845	Standard Cartridge Holder, 20 mm	ea



2.6 µm Minibore Columns (mm)						SecurityGuard ULTRA Cartridges‡	
Phases	30 x 2.1	50 x 2.1	75 x 2.1	100 x 2.1	150 x 2.1	3/pk	for 2.1 mm ID
EVO C18	00A-4725-AN	00B-4725-AN	—	00D-4725-AN	00F-4725-AN	AJ0-9298	for 2.1 mm ID



2.6 µm MidBore™ Columns (mm)						SecurityGuard ULTRA Cartridges‡	
Phases	30 x 3.0	50 x 3.0	75 x 3.0	100 x 3.0	150 x 3.0	3/pk	for 3.0 mm ID
EVO C18	00A-4725-Y0	00B-4725-Y0	—	00D-4725-Y0	00F-4725-Y0	AJ0-9297	for 3.0 mm ID



5 µm Minibore Columns (mm)						SecurityGuard ULTRA Cartridges‡	
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk	for 2.1 mm ID	
EVO C18	00A-4633-AN	00B-4633-AN	00D-4633-AN	00F-4633-AN	AJ0-9298	for 2.1 mm ID	

for 2.1 mm ID

5 µm MidBore™ Columns (mm)						SecurityGuard ULTRA Cartridges‡	
Phases	30 x 3.0	50 x 3.0	100 x 3.0	150 x 3.0	3/pk	for 3.0 mm ID	
EVO C18	00A-4633-Y0	00B-4633-Y0	00D-4633-Y0	00F-4633-Y0	AJ0-9297	for 3.0 mm ID	

for 3.0 mm ID

5 µm Analytical Columns (mm)						SecurityGuard ULTRA Cartridges‡	
Phases	50 x 4.6	100 x 4.6	150 x 4.6	250 x 4.6	3/pk	for 4.6 mm ID	
EVO C18	00B-4633-E0	00D-4633-E0	00F-4633-E0	00G-4633-E0	AJ0-9296	for 4.6 mm ID	

for 4.6 mm ID

\*SecurityGuard ULTRA Cartridges require holder,

Part No.: AJ0-9000

\*\*SemiPrep SecurityGuard Cartridges require holder,

Part No.: AJ0-9281

\*PREP SecurityGuard Cartridges require holder,

Part No.: AJ0-8223

# Luna™ One of The World's Leading LC Columns



## Luna C18

### Ordering Information

5 µm MidBore and Analytical Columns (mm)								SecurityGuard™ Cartridges (mm)	
Phases	30 x 3.0	50 x 3.0	150 x 3.0	250 x 3.0	30 x 4.6	50 x 4.6	75 x 4.6	4 x 2.0*	4 x 3.0*
C18(2)	00A-4252-Y0	00B-4252-Y0	00F-4252-Y0	00G-4252-Y0	00A-4252-E0	00B-4252-E0	00C-4252-E0	/10pk	/10pk

for ID: 2.0-3.0 mm 3.2-8.0 mm

5 µm Analytical and Semi-Prep Columns (mm)					SecurityGuard™ Cartridges (mm)	
Phases	100 x 4.6	150 x 4.6	250 x 4.6	250 x 10	4 x 3.0*	10 x 10†
C18(2)	00D-4252-E0	00F-4252-E0	00G-4252-E0	00G-4252-N0	AJ0-4287	AJ0-7221

for ID: 3.2-8.0 mm 9-16 mm

\*SecurityGuard™ Analytical Cartridges require holder, Part No.: [KJ0-4282](#)

†SemiPrep SecurityGuard™ Cartridges require holder, Part No.: [AJ0-9281](#)

## Luna Omega PS C18 and Luna C18

### Ordering Information

1.6 µm Microbore Columns (mm)				SecurityGuard™ ULTRA Cartridges‡	
Phases	50 x 1.0	100 x 1.0	150 x 1.0	150 x 2.1	3/pk
PS C18	—	<a href="#">00D-4752-A0</a>	—	<a href="#">00F-4752-AN</a>	<a href="#">AJ0-9508</a>
C18	<a href="#">00B-4742-A0</a>	<a href="#">00D-4742-A0</a>	<a href="#">00F-4742-A0</a>	<a href="#">00F-4742-AN</a>	<a href="#">AJ0-9502</a>
1.6 µm Minibore Columns (mm)				SecurityGuard™ ULTRA Cartridges‡	
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk
PS C18	<a href="#">00A-4752-AN</a>	<a href="#">00B-4752-AN</a>	<a href="#">00D-4752-AN</a>	<a href="#">00F-4752-AN</a>	<a href="#">AJ0-9508</a>
C18	<a href="#">00A-4742-AN</a>	<a href="#">00B-4742-AN</a>	<a href="#">00D-4742-AN</a>	<a href="#">00F-4742-AN</a>	<a href="#">AJ0-9502</a>

for 2.1 mm ID

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